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EFFECT OF SOME BIOSTIMULATING SUBSTANCES ON ZONAL GERANIUM GROWTH AND FLOWERING UNDER IRRIGATION WATER SALINITY

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ABSTRACT: This pots investigation was carried out during the two successive growing seasons of 2019 and 2020 at the Nursery of Ornamental plants, Horticulture Department, Faculty of Agriculture, Minia University, to examine the response of pink zonal geranium (Pelargonium × hortorum L.H. Bailey) plants, grown in sandy calcareous soil, to four levels of salinity and three treatments of biostimulants (seaweed extract, amino acid and control) as well as their interaction in terms of different vegetative growth parameters, root characters, flowering aspects and some chemical composition. The results pointed out that vegetative growth traits (plant height, number of branches and number of leaves/plant), root growth parameters (root fresh and dry weights), flowering aspects (number of inflorescences/plant and diameter of inflorescence) as well as photosynthetic pigments, N, P and K % were significantly decreased as a result to irrigation with saline water with the highest reductions were obtained with 0.75%. At the same time, salinity treatments increased Na, Ca and Cl % and proline content (mg/g f.w.). In addition, application of plants with any of the biostimulants (seaweed extract or amino acid, each at 200 ppm) led to a significant increase in all previous characteristics of vegetative growth, root growth, flowering aspects and some chemical composition (N, P, K and Ca %) compared to control. Therefore, biostimulant treatments decreased both Na and Cl % as well as proline content (mg/g f.w.). The combination treatments of tap water with amino acid (200 ppm) were superior than other interaction treatments for all studied parameters except in K %.

Key words: *Pelargonium* × *hortorum*, salinity, seaweed extract, amino acids, vegetative growth, chemical composition

INTRODUCTION

The zonal (cutting) geranium perennially ranks among the top five greenhouse flowering potted plants in total production units and whole value in the United States (Tayama and Carver, 1990). *Pelargonium* × *hortorum* L.H. Bailey, zonal geranium, belonging to the Family Geraniaceae are among the most economically important

bedding and pot plants. Currently, North America and Europe are the major producers and distributors of zonal pelargonium with global annual sales amounting to \$700 million (Mithila *et al.*, 2001).

About one-third of nearly 160 million hectares of cultured land under irrigation universally, is already salt-affected soils (Singh and Chatrath, 2001). Salinity stress seriously affected more than a third of the

world's cultivated lands (Bayat *et al.*, 2012), caused many harmful effects on plants, a sharp decrease in the yield of many plants and biomass, leads to growth reduction as well as metabolic changes (Abdelaal, 2015 and El-Banna and Abdelaal, 2018).

Seaweed extracts are an important source of plant growth regulators, containing amino acids, mineral nutrients, organic osmolites, vitamins, and vitamin precursors. In recent years, the use of natural seaweed extract as plant growth stimulation has allowed for its replacement in place of traditional synthetic fertilizers (Sahoo, 2000 and Khan *et al.*, 2009).

Amino acids are a well-known biostimulating material that has positive effects on plant growth, flowering, yield and significantly mitigates the adverse effects caused by abiotic stresses (Kowalczyk and Zielony, 2008).

Therefore, the aim of the present study was to evaluate the role of some biostimulants (seaweed extract and amino acid, each at 200 ppm) in reducing the harmful effects of irrigation water salinity on zonal geranium (*Pelargonium* × *hortorum* L.H. Bailey) plants under Minia city conditions.

MATERIALS AND METHODS

A pot investigation was carried out during the two successive growing seasons

of 2019 and 2020 at the Nursery of Ornamental plants, Horticulture Department, Faculty of Agriculture, Minia University, Cutting of pelargonium zonal Egypt. (Pelargonium × hortorum L.H. Bailey) were obtained from the Nursery. Average cut length was 12.0 cm and the cutting diameter 0.6 cm, having 3 leaves. experimental was arranged in a randomized complete block design (RBCD) in a split plot design with three replicates, each replicate containing 6 cuttings. The main plot (A) includes four levels of irrigation water salinity (0.25, 0.50 and 0.75%, equal 2.5, 5.0 and 7.5 g/l of both NaCl + CaCl₂ w/w)), while three treatments of biostimulants (seaweed extract, amino acid, each at 200 ppm, and control) occupy the sub plot. Therefore, the experiment consists of 12 treatments. The physical and chemical analyses of the used soil were presented in Table (a).

product contains seaweed Algeser extract, which was released from Shoura Chemical Company, Cairo-Alexandria Desert Road, Giza Governorate, Egypt. The chemical properties of the seaweed extract used in the experiment in both seasons were presented in Table (b). Also, Aminoactal product contains a blend of three amino acids (methionine, treptophan and cysteine), also, was obtained from Shoura Chemical Company.

Table a. Physical and chemical analysis of the used soil during the two seasons of 2018/2019 and 2019/2020.

2010/20	1) and 201)	12020.						
Soil characters	Val	lues	G-91 -1	7	Values			
	2018/2019	2019/2020	Soil characters	2018/2019	2019/2020			
Physical properties	:		Exchangeable nutrients	s (mg/100 g soil):				
Sand (%)	90.0	91.0	Ca++	8.21	8.43			
Silt (%)	7.30	6.40	$\mathbf{M}\mathbf{g}^{\scriptscriptstyle{++}}$	1.53	1.59			
Clay (%)	2.70	2.60	Na ⁺	0.99	1.05			
Soil type	Sandy	Sandy	\mathbf{K}^{+}	0.92	1.03			
Chemical propertie	es:		DTPA-Extractable nutr	rients:				
pH (1:2.5 paste)	8.15	8.25	Fe (ppm)	1.12	1.28			
E.C. (dS/m)	1.07	1.10	Cu (ppm)	0.40	0.44			
O.M. (%)	0.05	0.04	Zn (ppm)	0.35	0.29			
CaCO ₃ (%)	14.88	14.90	Mn (ppm)	0.55	0.66			

2020.			
Character	Values	Character	Values
Moisture (%)	6.0	S (%)	3 – 9
O.M. (%)	45 - 60	Mg (%)	0.5 - 0.9
Inorganic matter (%)	45 - 60	Cu (ppm)	1.0 - 6.0
Protein (%)	6 - 8	Fe (ppm)	50 - 200
Carbohydrate (%)	35 - 50	Mn (ppm)	5 - 12
Aliginic acid (%)	10 - 20	Zn (ppm)	10 - 100
Mannitol (%)	4 - 7	B (ppm)	20 - 100
Total N (%)	1.0 - 1.5	Mo (ppm)	1 - 5
P (%)	0.02-0.09	Cytokinins (%)	0.02
K (%)	1.0 - 1.2	IAA (%)	0.03
Ca (%)	0.2 - 1.5	ABA (%)	0.01

Table b. Chemical properties of seaweeds extract used in both seasons of 2019 and 2020.

Cuttings were planted in the last week of February 2019 and 2020 in plastic pots (20 cm diameter). Each was filled with 4.0 kg sandy calcareous soil in the Nursery of Ornamental plants, each contained 3 cuttings seedlings were thinned seedling/pot after one month from planting date (last week of March). The irrigation water salinity started on the first day of April and was repeated every two days with 300 ml/pot tap water or saline solution and continued until the end of the experiment (last week of September). Both seaweed extract and amino acid were applied 5 times at one-month intervals, starting from 7th April.

Data recorded:

Vegetative growth traits (plant height, number of branches and number of leaves/plant), root growth parameters (root fresh and dry weights), flowering aspects (number of inflorescences/plant and diameter of inflorescence) as well as some chemical composition [photosynthetic pigments and proline content (mg/g f.w), N, P, K, Ca, Na and Cl (%)].

N, P, K, Ca, Na and Cl content of leaves were done as described by ICARDA (2013). Chlorophyll content in fresh leaves samples (mg/g. f.w.) according to Fadl and Sari El–Deen (1978). Proline content was determined according to Bates *et al.* (1973) method.

The obtained results were tabulated and statistically analyzed according to MSTAT-

C (1986), and LSD test at 5% was followed to compare the means of treatments.

RESULTS

1. Vegetative growth traits:

Data presented in Table (1) showed that all vegetative growth parameters of zonal pelargonium (plant height, number of branches and number of leaves/plant) were significantly decreased by using the salinity treatments (0.25, 0.50 and 0.75%) as compared to the control in both seasons. It could be noticed that the greatest vegetative growth was obtained with control, while the lowest vegetative growth was recorded with the high irrigation water salinity level (0.75%).

These results were in agreement with those recorded by Valdés et al. (2015) and Breś et al. (2017) on Pelargonium × hortorum; Fornes et al. (2007) on Petunia hybrid, Cassaniti et al. (2009) on Grevillea juniperina, Khalid et al. (2014) on Nigella sativa; Rehman et al. (2014) on Kardinal (Rose), García-Caparrós et al. (2016) on Aloe vera, Ahir et al. (2017) on Tuberose, Francisco et al. (2017) on Catharanthus roseus, Shahmoradi and Naderi (2018) on winter jasmine, and Gadallah et al. (2020) and Singh et al. (2021) on Pelargonium graveolens.

Regarding the impact of biostimulants treatments on vegetative growth traits, the two used treatments of biostimulants (seaweed extract and amino acid, each at 200

Table 1. Effect of irrigation water salinity and some biostimulants treatments on plant height, number of branches and number of leaves/plant of zonal geranium plants during the first and second seasons.

plants daring	Irrigation water salinity levels (%) (A)										
Biostimulants treatments (B)	0.0	0.25	0.50	0.75	Mean (B)	0.0	0.25	0.50	0.75	Mean (B)	
		The 1	^{id} season	season (2020)							
	Plant height (cm)										
Control	23.00	21.00	13.00	11.00	17.00	23.46	21.42	13.26	11.22	17.34	
Seaweed extract (200 ppm)	25.30	24.79	23.83	22.81	24.18	25.81	25.29	24.31	23.27	24.67	
Amino acid (200 ppm)	26.56	26.01	24.99	23.74	25.33	27.09	26.53	25.49	24.21	25.83	
Mean (A)	24.85	23.93	20.61	19.18		25.45	24.41	21.02	19.57		
L.S.D. at 5%	A: 0.93		B: 0.28	AB: 0.56		A: 1.02		B: 0.29	AB	3: 0.58	
				Numb	er of br	anches/j	plant				
Control	4.00	2.67	2.60	2.23	2.88	4.08	2.72	2.65	2.28	2.93	
Seaweed extract (200 ppm)	4.67	3.33	3.00	2.33	3.33	4.76	3.40	3.06	2.38	3.40	
Amino acid (200 ppm)	6.00	4.66	3.33	2.67	4.17	6.12	4.75	3.40	2.72	4.25	
Mean (A)	4.89	3.55	2.98	2.41		4.99	3.62	3.04	2.46		
L.S.D. at 5%	A: 1.	34	B: 0.70	AB	3: 1.40	A: 1.	39	B: 0.71	AB	3: 1.48	
				Nun	iber of l	eaves/pl	ant				
Control	14.33	12.00	9.33	7.00	10.67	14.41	12.20	9.53	7.68	10.96	
Seaweed extract (200 ppm)	15.00	14.30	12.20	9.10	12.65	14.86	14.00	12.00	9.00	12.49	
Amino acid (200 ppm)	16.80	16.00	13.00	11.33	14.28	17.00	16.11	13.31	11.46	14.47	
Mean (A)	15.80	14.10	11.51	9.14		15.46	14.10	11.62	9.38		
L.S.D at 5%	A: 1.	20	B: 0.40	B: 0.40 AB: 0.5		A: 1.30		B: 0.34	AB	3: 0.68	

ppm) significantly increased all studied vegetative growth parameters in both seasons compared to the control. The treatment of amino acid was more effective than seaweed extract in both seasons.

Many researchers proved the enhancement effect of biostimulants on vegetative growth characters, such as Krajnc et al. (2012) on zonal pelargonium. Youssef (1997) on Delphinium ajacis, Thirumaran et al. (2009a), on Abelmoschus esculentus, Thirumaran et al. (2009b) on cluster bean, Akila and Jeyadoss (2010) on Helianthus annus, Sridhar and Rengasamy (2010) and Nofal et al. (2021) on Tagetes erecta, Abd El-Aziz et al. (2011) and Kumareswari and Rani (2015) on Amaranthus, De Luncia and Vecchietti (2012), on Lilium, Talib et al. (2012) and Nofal et al. (2015) on Calendula officinalis, Hassan (2016) and Khalil (2021) on Gladiolus grandiflorus, and Roshani and Asadi-Gharneh (2019) on Pelargonium graveolens, concerning the important role of seaweed extract.

In addition, Hassn *et al.* (2015) on *Pelargonium* × *hortorum*, Abd El-Aziz *et al.*

(2009), Hassan (2016) and Khalil (2021) on gladiolus, De lunica and Vecchietti (2012) and El-Naggar *et al.* (2013) on *lilium*, Nazmy and Arafa (2017) on *Calendula officinalis*, Nosir and El-Kinany (2019) on *Rosa hybrida*, Roshani and Asadi-Gharneh (2019) on *Pelargonium graveolens*, and El-Kinany (2020) on pansy plants, concerning the effect of amino acids.

The interaction between main and subplots treatments was significant for plant height, number of branches and number of leaves/plant in both seasons. Irrigated plants with tap water or low salinity level (0.25 %) in combination with amino acid led to produce the greatest values.

2. Root growth parameters:

Data presented in Table (2) clarified that irrigation water salinity treatments (0.25, 0.50 and 0.75%) had negative effects on fresh and dry weights of roots in both seasons compared with the control. The reduction in the root fresh weight due to 0.75% was 11.31% in both seasons compared with the control. While 0.75% salinity level declined root dry weight by

Table 2. Effect of irrigation water salinity and some biostimulants treatments on root growth and flowering parameters of zonal geranium plants during the first and second seasons.

and second sec	Irrigation water salinity levels (%) (A)										
Biostimulants treatments (B)	0.0	0.25	0.50	0.75	Mean (B)	0.0	0.25	0.50	0.75	Mean (B)	
		The 1s	^t season (^d season	(2020)		
	Root fresh weight (g)										
Control	18.24	16.80	16.44	15.60	18.24	20.06	18.48	18.08	17.16	18.45	
Seaweed extract (200 ppm)	20.28	19.80	19.32	18.24	20.28	22.31	21.78	21.25	20.06	21.35	
Amino acid (200 ppm)	21.96	21.48	21.00	19.80	21.96	24.16	23.63	23.10	21.78	23.17	
Mean (A)	20.16	19.36	18.92	17.88	20.16	22.18	21.30	20.81	19.67		
L.S.D. at 5%	A: 0.	82	B: 0.25	AB	3: 0.50	A: 0	.90	B: 0.27	AE	3: 0.54	
				Ro	ot dry v	veight (g	g)				
Control	5.47	5.04	4.93	4.68	5.03	6.02	5.54	5.42	5.15	5.53	
Seaweed extract (200 ppm)	6.08	5.94	5.80	5.47	5.82	6.69	6.53	6.38	6.02	6.41	
Amino acid (200 ppm)	6.59	6.44	6.30	5.94	6.32	7.25	7.09	6.93	6.53	6.95	
Mean (A)	6.05	5.81	5.68	5.36		6.65	6.39	6.24	5.91		
L.S.D. at 5%	A: 0.	26	B: 0.10	AB	3: 0.20	A: 0	.30	B: 0.15	AE	3: 0.30	
			N	umber	of inflo	rescence	es/plant				
Control	6.00	4.00	3.00	1.00	3.50	6.03	4.20	3.15	1.05	3.61	
Seaweed extract (200 ppm)	6.10	5.93	5.70	5.47	5.80	6.41	6.23	5.99	5.74	6.09	
Amino acid (200 ppm)	7.32	7.17	7.03	6.75	7.07	7.69	7.53	7.38	7.09	7.42	
Mean (A)	6.47	5.70	5.24	4.41		6.71	5.99	5.51	4.63		
L.S.D at 5%	A: 0.	78	B: 0.09	AB	B: 0.18 A: 0		.74	B: 0.11	AE	3: 0.22	
			I	Diamete	er of infl	lorescen	ce (cm)				
Control	7.40	7.00	6.00	4.00	6.10	7.70	7.28	6.24	4.16	6.35	
Seaweed extract (200 ppm)	7.77	7.35	6.30	4.20	6.41	8.09	7.64	6.55	4.37	6.66	
Amino acid (200 ppm)	7.84	7.69	7.53	7.22	7.57	8.15	7.99	7.83	7.51	7.87	
Mean (A)	7.67	7.35	6.61	5.14		7.98	7.64	6.78	5.35		
L.S.D at 5%	A: 0.	34			B: 0.16 A: 0		.36	B: 0.09	AE	3: 0.18	

11.41 in the first season, and by 11.13% in the second one comparing to the control (16.80) in the first season, and 17.75, 17.34 and 16.39 cm in the second season compared with the control (18.48), respectively. The other two salinity treatments (0.25 and 0.50%) gave an intermediate values.

Similar results were recorded by Bahadoran and Salehi (2015) on *Polianthes tuberosa* and Ahmed (2017) on Rose China and Raslan.

Regarding the impact of the biostimulants treatments, data listed in Table (2) indicated that using either seaweed extract or amino acid, each at 200 ppm, significantly improved root fresh and dry weights compared with control in both seasons. Amino acid treatment was superior

to seaweed extract in this concern in both seasons. Such superior treatment increased root dry weight by 8.59% in the first season and 8.42% in the second season compared to seaweed extract, respectively.

The enhancement influence of seaweed extract on root fresh and dry weights were reported by Krajnc *et al.* (2012) on zonal geranium Thirumaran *et al.* (2009a) on *Abelmoschus esculentus*, Thirumaran *et al.* (2009b) cluster bean and Osman and Salem (2011) on sunflower, concerning the effect of seaweed extract. Whereas, Shehata *et al.* (2011) on celeriac plants and Roshani and Asadi-Gharneh (2019) on *Pelargonium graveolens*, regard to the impact of amino acid.

The interaction between the main and sub-plots treatments $(A \times B)$ was significant for root fresh and dry weights in both seasons. Planting pelargonium zonal in sandy calcareous soil, irrigated with unsalinized water or low salinity water (0.25%) and sprayed with amino acid (200 ppm) produced the heaviest root fresh and dry weights as clearly shown in Table (2).

3. Flowering parameters:

Data in Table (2) indicated that number of inflorescence/plant and diameter of inflorescence were reduced by increasing salinity level, till 0.50 and 0.75% salinity levels significantly decreased both characters in two growing seasons compared to the control.

Similarly, the same trends were obtained by Grieve and Poss (2010) on ornamental sunflower, Ahmad *et al.* (2013) on *Rosa hybrid*, Chauhan and Ambast (2014) on marigold, Bahadoran and Salehi (2015) on *Polianthes tuberosa* and Ahmed (2017) *Hibiscus rosa-sinensis*.

Data presented in Table (2) showed that number of inflorescence/plant and diameter of zonal geranium inflorescence was significantly increased by using seaweed extract or amino acid, each at 200 ppm, compared to the control in both seasons. Moreover, amino acid treatment was more effective than seaweed extract in this concern.

Many authors proved the augmented effects of seaweed extract on diameter of inflorescence such as Abd El-Aziz *et al.* (2011) on *Amaranthus tricolor* and Talib *et al.* (2012) on *Calendula officinalis*. Also, the improvement roles of amino acid on diameter of inflorescence were recorded by Afifipour and Kosh-Khui (2015) on *Polianthes tuberose* and Mahdi and Saeed (2019) on gerbera plants.

The interaction between the two experimental factors $(A \times B)$ was significant for number of inflorescence/plant and diameter of inflorescence in both seasons. The best interaction treatments were to

plants irrigated with tap water or a low level of salinity (0.25%) and plants sprayed with amino acid at 200 ppm.

4. Chemical composition:

a. Photosynthetic pigments:

Data presented in Tables (3) indicated that all used salinity treatments (0.25, 0.50 and 0.75%) significantly decreased chlorophyll a, b and carotenoids as compared to control in both seasons. The treatment of 0.75% saline water resulted the greatest reduction in chlorophyll a (2.467 and 2.491 mg/g f.w.), chlorophyll b (0.782 and 0.788 mg/g f.w.) and carotenoids (0.813 and 0.829 mg/g f.w.) in the first season. While the low level of salinity (0.25%) slightly decreased photosynthetic pigments. The same trend was obtained in the second season.

Similar results were obtained by Valdés et al. (2015), Breś et al. (2016) and Breś et al. (2017) on Pelargonium × hortorum, Garnett et al. (2002), Khetsha and Sedibe (2015) and Singh et al. (2021) on Pelargonium graveolens, Khalid et al. (2010), Al-Khafajy et al. (2020) and El-Shawa et al. (2020) on Calendula officinalis, Bahadoran and Salehi (2015) and Ahir et al. (2017) on Polianthes tuberosa, Ahmed (2017) on Rose China and Shahmoradi and Naderi (2018) on winter jasmine.

Regarding the effect of biostimulants (seaweed extract and amino acid, each at 200 ppm), using each of them significantly increased chlorophyll a, b and carotenoids as compared with control (Table, 3). Moreover, amino acid treatment was more effective in this concern.

Several researchers demonstrated the positive impact of seaweed extract on photosynthetic pigments such as Krajnc *et al.* (2012) on *Pelargonium* × *hortorum*, Blunden *et al.* (1997), on some plant species, Thirumaran *et al.* (2009a) on *Abelmoschus esculentus*, Thirumaran *et al.* (2009b) on cluster bean, Sridhar and Rengasamy (2010) on *Tagetes erecta*, and Akila and Jeyadoss (2010), on *Helianthus annus*, Abd El-Aziz *et al.* (2011) and Kumareswari and Rani (2015)

Table 3. Effect of irrigation water salinity and some biostimulants treatments on photosynthetic pigments (mg/g f.w.) of zonal geranium plants during the first and second seasons.

una secona se			Irri	gation v	vater sal	linity lev	vels (%) (A)		
Biostimulants treatments (B)	0.0	0.25	0.50	0.75	Mean (B)	0.0	0.25	0.50	0.75	Mean (B)
		The 1	st season	(2019)						
				Chlo	rophyll	a (mg/g	f.w.)			
Control	2.358	2.003	1.946	1.947	1.989	2.382	2.023	1.966	1.663	2.009
Seaweed extract (200 ppm)	3.655	2.494	2.428	2.414	2.748	3.692	2.519	2.452	2.438	2.775
Amino acid (200 ppm)	3.983	3.807	3.558	3.340	3.672	4.023	3.845	3.593	3.373	3.709
Mean (A)	3.332	2.768	2.644	2.467		3.366	2.796	2.670	2.491	
L.S.D. at 5%	A: 0.088		B: 0.005 AB: N.S.		3: N.S.	A: 0.083		B: 0.007	AB	: N.S.
	Chlorophyll b (mg/g f.w.)									
Control	0.766	0.648	0.629	0.529	0.643	0.774	0.654	0.635	0.534	0.649
Seaweed extract (200 ppm)	1.201	0.811	0.779	0.785	0.894	1.213	0.819	0.787	0.793	0.903
Amino acid (200 ppm)	1.307	1.249	1.166	1.033	1.189	1.320	1.261	1.178	1.043	1.201
Mean (A)	1.091	0.903	0.858	0.782		1.102	0.911	0.869	0.788	
L.S.D. at 5%	A: 0.0	062	B: 0.011	AB	3: N.S.	A: 0.0	066	B: 0.013	AB	: N.S.
				Car	otenoids	(mg/g f	.w.)			
Control	0.806	0.688	0.668	0.569	0.683	0.822	0.702	0.681	0.580	0.696
Seaweed extract (200 ppm)	1.252	0.851	0.829	0.826	0.939	1.276	0.868	0.846	0.843	0.958
Amino acid (200 ppm)	1.346	1.289	1.206	1.043	1.222	1.375	1.315	1.230	1.064	1.246
Mean (A)	1.135	0.943	0.901	0.813		1.158	0.962	0.919	0.829	
L.S.D at 5%	A: 0.0)92			B: N.S. A: 0.081		081	B: 0.043 AB		: N.S.

on Amaranthus, Osman and Salem (2011) on sunflower, De Luncia and Vecchietti (2012), on Lilium, Talib et al. (2012) on Calendula officinalis and Hassan (2016) and Khalil (2021) on Gladiolus grandiflorus.

Furthermore, the enhancement effects of amino acid on augmented photosynthetic pigments were reported by Abd El-Aziz et al. (2009), Sewedan et al. (2012), Hassan (2016) and Khalil (2021) on gladiolus, Ibrahim et al. (2010) on Helichrysum bracteatum, Mahgoub et al. (2011) on Dahila pinnata, Rahmatzadeh et al. (2012) on Catharanthus roseus, Ali and Hassan (2013) on Tagetes erecta, Aly et al. (2014) on Kapok, Rafiee et al. (2013) and Nazmy and Arafa (2017) on Calendula officinalis and Roshani and Asadi-Gharneh (2019) on Pelargonium graveolens.

The interaction between the main and sub-plots (A x B) was not significant for chlorophyll a, b and carotenoids in both seasons as clearly shown in Table (3).

b. N, P, K and Ca (%):

Data presented in Tables (4) showed that all levels of salinity treatments (0.25, 0.50 and 0.75%) significantly decreased N, P and K percentages in pelargonium leaves, however, they increased Ca (%) as compared to control in both seasons. The highest percentages of N, P and K in the first season reached 2.80, 0.42 and 1.98%, facing the lowest percentages, 2.02, 0.31 and 1.44%, respectively. Similarly, in the second season, the highest values were 2.94, 2.08 and 2.12%, while, the lowest values, 2.12, 0.33 and 1.51%, respectively. For N, P and K%, the highest values were recorded with control treatments in both seasons, while the lowest values were produced with the highest level of saline irrigation water (0.75%). Contrarily, the highest values of Ca % were obtained with the highest level of saline irrigation water as it gave 1.64% and 1.72% in the first and second seasons, respectively.

Table 4. Effect of irrigation water salinity and some biostimulants treatments on N, P, K and Ca (%) of zonal geranium plants during the first and second seasons.

Irrigation water salinity levels (%) (A)													
			Irrig	gation v	water sal	linity le	vels (%) (A)					
Biostimulants treatments (B)	0.0	0.25	0.50	0.75	Mean (B)	0.0	0.25	0.50	0.75	Mean (B)			
		The 1 st season (2019)						The 2 nd season (2020)					
		Nitrogen (%)											
Control	2.34	2.11	1.86	1.67	2.00	2.46	2.22	1.95	1.75	2.10			
Seaweed extract (200 ppm)	2.89	2.63	2.33	2.09	2.49	3.03	2.76	2.45	2.19	2.61			
Amino acid (200 ppm)	3.18	2.89	2.56	2.30	2.73	3.34	3.03	2.69	2.42	2.87			
Mean (A)	2.80	2.54	2.25	2.02		2.94	2.67	2.36	2.12				
L.S.D. at 5%	A: 0.	20	B: 0.15	AE	3: 0.30	A: 0.	22	B: 0.18	AE	3: 0.36			
					Phospho	rus (%)						
Control	0.37	0.35	0.32	0.27	0.33	0.40	0.37	0.34	0.28	0.35			
Seaweed extract (200 ppm)	0.43	0.41	0.37	0.31	0.38	0.45	0.43	0.39	0.33	0.40			
Amino acid (200 ppm)	0.47	0.45	0.41	0.35	0.42	0.49	0.47	0.43	0.37	0.44			
Mean (A)	0.42	0.40	0.37	0.31		0.44	0.42	0.39	0.33				
L.S.D. at 5%	A: 0.	01	B: 0.02	B: 0.02 AB: 0.04		A: 0.01		B: 0.02	AE	3: 0.04			
					Potassiu	ım (%)							
Control	1.83	1.74	1.57	1.33	1.62	1.92	1.84	1.65	1.40	1.70			
Seaweed extract (200 ppm)	2.01	2.00	1.80	1.53	1.84	2.21	2.12	1.89	1.61	1.96			
Amino acid (200 ppm)	1.20	1.91	1.72	1.46	1.57	2.11	2.02	1.81	1.53	1.87			
Mean (A)	1.98	1.88	1.70	1.44		2.08	1.99	1.78	1.51				
L.S.D at 5%	A: 0.	03	B: 0.05	AE	B: 0.10	A: 0.	04	B: 0.06	AE	B: 0.12			
					Calciu	m (%)							
Control	1.13	1.19	1.31	1.51	1.29	1.19	1.25	1.38	1.59	1.35			
Seaweed extract (200 ppm)	1.24	1.31	1.44	1.66	1.41	1.30	1.38	1.52	1.74	1.48			
Amino acid (200 ppm)	1.30	1.38	1.51	1.74	1.48	1.37	1.45	1.59	1.83	1.56			
Mean (A)	1.22	1.29	1.42	1.64		1.28	1.36	1.49	1.72				
L.S.D at 5%	A: 0.	06	B: 0.04			A: 0.08		B: 0.05	AB: 0.10				

The adverse effects of salinity on NPK were obtained by Cassaniti *et al.* (2009) on *Grevillea juniperina*, Leithy *et al.* (2009) and Khetsha and Sedibe (2015) and Singh *et al.* (2021) on *Pelargonium graveolens*, and Ahmed (2017) on Rose China.

Concerning the impact of biostimulants, data listed in Table (4) clarified that the two treatments of seaweed extract and amino acid, each at 200 ppm, significantly increased NPK and Ca % in both seasons as compared with control. Moreover, amino acid treatment was superior than seaweed extract in increasing the N, P and Ca percentages. On the other hand, seaweed extract was more effective in increasing potassium percentage in pelargonium leaves.

Several researchers pointed out the improvement role of biostimulants on N, P, K and Ca percentages such as Abd El-Aziz *et al.* (2011) on *Amaranthus*, Hassan (2016)

and Khalil (2021) on *Gladiolus grandiflorus*, regard to seaweed extract.

Furthermore, the enhancement effects of amino acid on augmented N, P and K percentages were reported by Abd El-Aziz *et al.* (2009), Sewedan *et al.* (2012), Hassan (2016) and Khalil (2021) on gladiolus, Nofal *et al.* (2021) on African marigold.

The interaction between the main and sub-plots ($A \times B$) was significant for N, P, K and Ca percentages in both seasons as clearly shown in Table (4). The highest values of N, P and K percentages were recorded with tap water in combination with amino acid or seaweed extract, each at 200 ppm or irrigated plants with low level of salinity (0.25%) plus amino acid. However, the highest value of Ca percentage was obtained with the highest level of salinity (0.75%) in combination with amino acid at 200 ppm.

c. Na, Cl and proline content:

Data presented in Table (5) proved that unlike the trend of N, P and K%, sodium, chloride and proline contents were significant increase by using any of the three levels of irrigation water salinity (0.25, 0.50 and 0.75%) in pelargonium leaves as compared to control in both seasons.

Similar results were recorded by Valdés et al. (2015), Breś et al. (2016) and Breś et al. (2017) on zonal geranium, Prasad et al. (2006) and Leithy et al. (2009) on Pelargonium graveolens, Fornes et al. (2007) on Petunia hybrida, Cassaniti et al. (2009) on Grevillea juniperina, Bahadoran and Salehi (2015) on tuberose Ahmed (2017) on Rose China and García-Caparrós and Lao (2018) on some ornamental plants.

Data presented in Table (5) indicated that spraying plants with biostimulants (seaweed extract and amino acid, each at 200 ppm) significantly decreased both of Na, Cl and proline content as compared to control in both seasons.

The interaction between the main and

sub-plots (A \times B) was significant for Na, Cl and proline content in both seasons as clearly shown in Table (5). The highest values in the first season (2.13, 1.78%) and 0.497 mg/g f.w., respectively) were recorded with the highest level of salinity (0.75%) without the addition of either amino acid or seaweed extract. Although, the lowest values of Na, Cl and proline in the first season (1.49%, 1.27%) and 0.363 mg/g f.w. were obtained with the lowest level of salinity (0.25 %) in combination with amino acid at 200 ppm. The same trend was observed in the second season.

DISCUSSION

Treated plants with saline irrigation water (0.25, 0.50 and 0.75 %) led to a decrease in all vegetative growth traits, root growth parameters, flowering aspects and photosynthetic pigments as well as NPK %. On the other hand, Na, Ca, Cl and proline contents were increased. These results reflected the negative role of salinity which were explained by many authors as follows:

Reclaimed wastewater may contain toxic

Table 5. Effect of irrigation water salinity and some biostimulants treatments on sodium, chloride, and proline content of zonal geranium leaves during the first and second seasons.

	Irrigation water salinity levels (%) (A)										
Biostimulants treatments (B)	0.0	0.25	0.50	0.75	Mean (B)	0.0	0.25	0.50	0.75	Mean (B)	
		The 15	st season	(2019)	The 2 nd season (2020)						
					Sodiur	n (%)					
Control	1.64	1.73	1.92	2.13	1.86	1.72	1.82	2.02	2.24	1.95	
Seaweed extract (200 ppm)	1.56	1.64	1.82	2.02	1.76	1.65	1.72	1.91	2.12	1.85	
Amino acid (200 ppm)	1.49	1.57	1.75	1.94	1.69	1.57	1.65	1.83	2.03	1.77	
Mean (A)	1.57	1.65	1.83	2.03		1.65	1.73	1.92	2.13		
L.S.D. at 5%	A: 0.	07	B: 0.04	AB: 0.08		A: 0.06		B: 0.05	AE	3: 0.10	
	Chloride (%)										
Control	1.39	1.50	1.62	1.78	1.57	1.45	1.56	1.68	1.85	1.64	
Seaweed extract (200 ppm)	1.32	1.43	1.54	1.69	1.50	1.37	1.49	1.60	1.76	1.56	
Amino acid (200 ppm)	1.27	1.37	1.48	1.63	1.55	1.32	1.42	1.54	1.70	1.50	
Mean (A)	1.33	1.43	1.55	1.70		1.38	1.49	1.61	1.77		
L.S.D. at 5%	A: 0.	09	B: 0.04	AE	3: 0.08	A: 0.	07	B: 0.03	AE	3: 0.06	
				Proli	ne conte	nt (mg/	g f.w.)				
Control	0.382	0.402	0.447	0.497	0.432	0.397	0.418	0.465	0.517	0.449	
Seaweed extract (200 ppm)	0.374	0.393	0.438	0.487	0.423	0.389	0.409	0.456	0.506	0.440	
Amino acid (200 ppm)	0.363	0.381	0.425	0.472	0.410	0.378	0.396	0.442	0.491	0.427	
Mean (A)	0.373	0.392	0.437	0.485		0.388	0.408	0.454	0.505		
L.S.D at 5%	A: 0.0)15	B: 0.08	0.08 AB: 0.016		A: 0.018		B: 0.08	AB	: 0.016	

ions and high concentrations of salt, a problem whose magnitude will depend on the location of any wastewater treatment plant (Bañón *et al.*, 2011).

- Salinity caused a disturbance in water relationship resulting in a loss of turgor, inhibition of cell elongation, stomatal closure and a decrease in the intensity of photosynthesis (Munns and Tester 2008; Cassaniti et al., 2012).
- One of the first responses of plants to salinity is a decreased rate of leaf growth (Blum, 1986) primarily due to the osmotic effect of salt around the roots, which leads to a reduction in water supply to leaf cells. High external salt concentrations can also inhibit root growth (Wild, 1988), with a reduction in length and mass of roots (Shannon and Grieve, 1999) and of function.

Effect of seaweed extract:

The observed enhancement effect of seaweed extract in this study reflected physiological and biological roles and function of this stimulant material, which were explained by many researchers, that these extracts involved macro and microelements, amino acids and growth regulators (cytokinins, auxin and gibberellins), fatty acids and vitamins (Chapman and Chapman, 1980; Kingman and Moore, 1982).

Effect of amino acid:

The improvement impact of amino acid physiological reflected the functional of amino acid which was noted by many authors. Some amino acids can affect plant growth and development through their influence on IAA and GA biosynthesis, and it play an important role in plant metabolism and protein assimilation which are necessary for cell formation, and consequently increase plant weights (Phillips, 1971; Russel, 1982 and Walter and Nawacki, 1987). Also, they buffering the biosynthesis of other organic compounds like, pigments, alkaloids, enzymes, co-enzymes, tryptophan, purine and pyrimidine bases (Goss, 1973; Kamar and Omar, 1987).

From the previous physiological and biological studies, it might be concluded that salt stress had negative effects on vegetative growth, root growth, flowering aspects and photosynthetic pigments as well as NPK %, application plants with stimulant substances (seaweed extract and amino acid) which were responsible for enhancing different physiological processes alleviate the adverse effects of salinity stress. Which reflected on stimulant various vegetative growth traits, root growth, aspects and some chemical flowering composition, i.e. photosynthetic pigments, N, P, K, Na, Ca and Cl and proline content of zonal geranium (Pelargonium x hortorum L.H. Bailey) plants.

REFERENCES

Abd El-Aziz, N.G.; Taha, L.S. and Ibrahim, S.M.M. (2009). Some studies on the effect of putrescine, ascorbic acid and thiamine on growth, flowering and some chemical constituents of Gladiolus plants at Nubaria. Ozean J. of Applied Sci., 2(2):169-179.

Abd El-Aziz, N.G.; Mahgoub, M.H. and Siam, H.S. (2011). Growth, flowering and chemical constituents performance of *Amaranthus tricolor* plants as influenced by seaweed (*Aschophyllum nodosum*) extract application under salt stress conditions. J. of Applied Sci. Res., 7(11):1472-1484.

Abdelaal, K.A.A. (2015). Effect of salicylic acid and abscisic acid on morphophysiological and anatomical characters of faba bean plants (*Vicia faba* L.) under drought stress. J. Plant Production, Mansoura Univ., Egypt, 6(11):1771-1788.

Afifipour, Z. and Kosh-Khui, M. (2015). Efficacy of spraying a mixture of amino acids on the physiological and morphological characteristics of tuberose

- (*Polianthes Tuberosa* L.). International J. of Hort. Sc., 2(2):199-204.
- Ahir, M.P.; Singh, A. and Patil, S.J. (2017). Response of different salinity levels on growth and yield of tuberose cv. Prajwal. International Journal of Chemical Studies, 5(6):2150-2152.
- Ahmad, I.; Khan, M.A.; Qasim, M.; Ahmad, R. and Tauseef, U. (2013). Growth, yield and quality of *Rosa hybrida* L. as influenced by NaCl salinity. Journal of Ornamental Plants (Journal of Ornamental and Horticultural Plants), 3(3):143-153.
- Ahmed, M.A. (2017). Effect of salinity (NaCl + CaCl₂) and some soil additions on growth, flowering and chemical composition of rose of china (*Hibiscus rosa-sinensis* L.) plant. Middle East Journal of Agriculture Research, 6(2):302-314.
- Akila, N. and Jeyadoss. T. (2010). The potential of seaweed liquid fertilizer on the growth and antioxidant enhancement of *Helianthus annuus* L. Oriental J. of Chemistry, 26(4):1353-1360.
- Ali, E.F. and Hassan, F.A.S. (2013). Impact of foliar application of commercial amino acids nutrition on the growth and flowering of *Tagetes erecta*, L. plants. J. of Applied Sci. Res., 9(1):652-657.
- Al-Khafajy, R.A.; AL-Taey, D.K.A. and AL-Mohammed, M.H.S. (2020). The impact of water quality, bio fertilizers and selenium spraying on some vegetative and flowering growth parameters of *Calendula officinalis* L. under salinity stress. Int. J. Agric. Stat. Sci., 16(1):1175-1180.
- Aly, M.K.A.; Badran, F.S.; Sayed, R.M. and Ahmed, R.E. (2014). Response of Kapok tree seedlings to some biostimulant Substances. Minia J. of Agric. Res. Develop., 2(34):155-164.
- Bahadoran, M. and Salehi, H. (2015). Growth and flowering of two tuberose (*Polianthes tuberosa* L.) cultivars under

- deficit irrigation by saline water. Journal of Agricultural Science and Technology, 17(2):415-426.
- Bañón, S.; Miralles, J.; Ochoa, J.; Franco, J.A. and Sánchez-Blanco, M.J. (2011). Effects of diluted and undiluted treated wastewater on the growth, physiological aspects and visual quality of potted lantana and polygala plants. Scientia Horticulturae, 129(4), 869-876.
- Bates, L.S.; Waldren, R.P. and Teare, J.D. (1973). Rapid determination of proline for water stress studies. Plant and Soil, 39:205-207.
- Bayat, H.; Alirezaie, M. and Neamati, H. (2012). Impact of exogenous salicylic acid on growth and ornamental characteristics of calendula (*Calendula officinalis* L.) under salinity stress. J. Stress Physiol. and Biochem., 8(1):258-267.
- Blum, A. (1986). Salinity resistance, In: Blum, A. (ed.), Plant Breeding for Stress Environments, CRC Press, Boca Raton, USA, pp. 1163-1169.
- Blunden, G.; Jenkins, T. and Liu, Y.W. (1997). Enhanced leaf chlorophyll levels in plants treated with seaweed extract. Journal of applied phycology, 8(6):535-543.
- Breś, W.; Bandurska, H.; Kupska, A.; Niedziela, J. and Frąszczak, B. (2016). Responses of pelargonium (*Pelargonium*× *hortorum* L.H. Bailey) to long-term salinity stress induced by treatment with different NaCl doses. Acta Physiologiae Plantarum, 38(1):1-11.
- Breś, W.; Kupska, A.; Kleiber, T.; Bosiacki, M. and Mieloszyk, E. (2017). Influence of sodium chloride on selected growth parameters and macronutrient content in pelargonium leaves. Journal of Elementology, 22(1):209-218.
- Cassaniti, C.; Leonardi, C. and Flowers, T.J. (2009). The effects of sodium chloride on ornamental shrubs. Scientia Horticulturae, 122(4):586-593.

- Cassaniti, C.; Romano, D. and Flowers, T.J. (2012). The response of ornamental plants to saline irrigation water. In: Garcia-Garizabal, I. (ed), Irrigation-Water Management, Pollution and Alternative Strategies, InTech, Croatia, pp. 131–139.
- Chapman, V.J. and Chapman, D.J. (1980). Seaweeds and Their Uses, 3rd Ed. Chapman and Hall, London, New York, 334 p.
- Chauhan, S.K. and Ambast, S.K. (2014). Effect of salinity levels and plant spacing on growth and flowering behaviour of marigold. Ann. Pl. and Soil Res, 16 (2): 125-127.
- De Luncia, B.D. and Vecchietti, L. (2012). Type of Bio-stimulant and application method effects on stem quality and root system growth in L. A. Lily. European J. of Hort. Sci., 77(1):10-15.
- El-Banna, M.F. and Abdelaal, K.A.A. (2018). Response of strawberry plants grown in the hydroponic system to pretreatment with H₂O₂ before exposure to salinity stress. J. Plant Production, Mansoura Univ., 9:989-1001.
- El-Kinany, R.G. (2020). The beneficial role of salicylic acid, triacontanol and δ-aminolevulinic acid on the growth, flowering and chemical composition of pansy (*Viola wittrockiana* Gams) under salt stress conditions. Hortscience Journal of Suez Canal University, 9(1):13-30.
- El-Naggar, A.A.M.; Amani, I.A. and El-Tony, F.H. (2013). Response of *Longiflorum* × *asiatic* hybrid lilium plant to foliar spray with some amino acids. Alex. J. Agric. Res., 58(3):197-208.
- El-Shawa, G.M.; Rashwan, E.M., and Abdelaal, K.A. (2020). Mitigating salt stress effects by exogenous application of proline and yeast extract on morphophysiological, biochemical and anatomical characters of calendula plants. Scientific Journal of Flowers and Ornamental Plants, 7(4):461-482.

- Fadl, M.S. and Sari El-Deen, S.A. (1978). Effect of N6-benzyl adenine on photosynthetic pigments and total soluble sugars of olive seedlings grown under saline conditions. Egypt. J. Hort., 6(2):169-183.
- Fornes, F.; Belda, R.M.; Carrión, C.; Noguera, V.; García-Agustín, P. and Abad, M. (2007). Pre-conditioning ornamental plants to drought by means of saline water irrigation as related to salinity tolerance. Scientia Horticulturae, 113(1):52-59.
- Francisco, O.F.I.; de Medeiros, W.J.; Lacerda, C.F.D.; Neves, A.L. and Oliveira, D.R. (2017). Saline water irrigation managements on growth of ornamental plants. Revista Brasileira de Engenharia Agrícolae Ambiental, 21:739-745.
- Gadallah, L.S.; El-Ghadban, M.E.M.; Soliman, A.S.; Mehasen, H.R.A. and Abbas, M.S. (2020). Effect of foliar application of antisalinity substances on geranium (*Pelargonium graveolens*) plants under salt stress. Journal of Animal and Plant Sciences, 30(3):723-729.
- García-Caparrós, P. and Lao, M.T. (2018). The effects of salt stress on ornamental plants and integrative cultivation practices. Scientia horticulturae, 240: 430-439.
- García-Caparrós, P.; Llanderal, A.; Pestana, M.; Correia, P.J. and Lao, M.T. (2016). Tolerance mechanisms of three potted ornamental plants grown under moderate salinity. Scientia Horticulturae, 201: 84-91.
- Garnett, M.R.; Murch, S.J.; KrishnaRaj, S.; Dixon, M.A. and Saxena, P.K. (2002). The rhizofiltration of sodium from hydroponic fluid using scented geraniums. Water, Air, and Soil Pollution, 140(1):343-365.

- Goss, J.A. (1973). Physiology of Plants and Their Cells; Pergamon Press, Inc., New York, USA, 457 p.
- Grieve, C.M. and Poss, J.A. (2010). Response of ornamental sunflower cultivars 'Sunbeam' and 'Moonbright' to irrigation with saline wastewaters. Journal of Plant Nutrition, 33:1579-1592.
- Hassan, A.A. (2016). Response of Gladiolus Plant to Some Fertilization and Antioxidant Treatments. Ph.D. Thesis, Fac. Agric., Minia Univ., Egypt, 215 p.
- Hassn, F.A.; Al-Tahir, Z.A.A. and Salih, A.N. (2015). Effect of same amino acids and coconut liquid on vegetative growth, flowering, and vdofileoil of geranium (*Pelargonium hortorum* L.). Muthanna Journal of Agricultural Science, 3(1):1-20.
- Ibrahim, S.M.M.; Taha, L.S. and Farahat, M.M. (2010). Influence of foliar application of peptone on growth, flowering and chemical composition of *Helichrysum bracteatum* plants under different irrigation intervals. Ozean J. of Applied Sci., 3(1):143-155.
- ICARDA (2013). Methods of soil, plant and water analysis: A manual for the West Asia and North Africa region, 3rd edition. International Center for Agricultural Research in the Dry Areas, Beirut, Lebanon, 243 p.
- Kamar, M.E. and Omar, A. (1987). Effect of nitrogen levels and spraying with animal-forte (amino acids salvation) on yield of cucumber and potatoes. J. Agric. Mansoura Univ., 12 (4): 900-907.
- Khalid, K.A. and da Silva, J.A.T. (2010). Yield, essential oil and pigment content of *Calendula officinalis* L. flower heads cultivated under salt stress conditions. Scientia Horticulturae, 126 (2): 297-305.
- Khalid, K.A.; Shedeed, M.R.; Salvador, R.T.; Soliven, R.L.; Balagan, E.J.Y. and Abes, N.S. (2014). Influence of kinetin

- on growth and biochemical accumulation in *Nigella sativa* L. plants grown under salinity stress conditions. Thai. J. Agric. Sci., 47:195-203.
- Khalil, A.R.M. (2021). Physiological Studies on Gladiolus Plant. Ph.D. Thesis, Fac. Agric., Minia Univ., Egypt, 146 p.
- Khan, W.; Rayirath, U.P.; Subramanian, S.; Jithesh, M.N.; Rayorath, P.; Hodges, D.M.; Critchley, A.T.; Craigie, J.S.; Norrie, J. and Prithiviraj, B. (2009). Seaweed extracts as biostimulants of plant growth and development. J. Plant Growth Regul., 28:386–399.
- Khetsha, Z.P. and Sedibe, M.M. (2015). Effect of NaCl and root-media selection on yield attributes, oil composition and mineral composition of rose geranium (*Pelargonium graveolens* L.). Proc. the 2nd International Conference on Agriculture and Forestry, India, 1:152-166.
- Kingman, A.R. and Moore, J. (1982). Isolation, purification and quantification of several growth regulating substance in *Ascophyllum nodosum* (*Phaeophyceae*). Botanica Marina, 25:149-153.
- Kowalczyk, K. and Zielony, T. (2008). Effect of Aminoplant and Asahi on Yield and Quality of Lettuce Grown on Rockwool. Proc. of the Biostimulators in Modern Agriculture Conf., 7-8 Febuary, Warsaw, Poland.
- Krajnc, A.U.; Ivanus, A.; Kristl, J. and Susek, A. (2012). Seaweed extract elicits the metabolic responses in leaves and enhances growth of *Pelargonium* cuttings. Eur. J. Hort. Sci., 77: 170-81.
- Kumareswari, T. and Rani, S.M.V. (2015). Utilization of seaweeds to enhance growth and nutritive status of *Amaranthus caudatus* L. International J. of Res. Studies in Biosciences, 3(3):9-15.
- Leithy, S.; Gaballah, M.S. and Gomaa, A.M. (2009). Associative impact of bio-and organic fertilizers on geranium plants grown under saline conditions.

- International Journal of Academic Research, 1(1):17-23.
- Mahdi, S.A. and Saeed, A.K.A.J.M. (2019). Effect of tryptophan and phenolalanine on some biochemical components and floral traits of gerbera (*Gerbera jamesonii* L.) cv. Great Smoky Mountains. Plant Archives, 19(1):1051-1056.
- Mahgoub, M.H.; El-Aziz, N.G.A. and Mazhar, A.M.A. (2011). Response of *Dahlia pinnata* L. plant to foliar spray with putrescine and thiamine on growth, flowering and photosynthetic pigments. American-Eurasian Journal of Agricultural and Environmental Sciences, 10 (5): 769-775.
- Mithila, J.; Murch, S.J.; KrishnaRaj, S. and Saxena, P.K. (2001). Recent advances in pelargonium in vitro regeneration systems. Plant Cell Tiss. Org. Cult., 67: 1–9.
- MSTAT-C, (1986). A microcomputer program for the design management and analysis of agronomic research experiments (Version 4), Michigan State Univ., USA.
- Munns, R. and Tester, M. (2008). Mechanisms of salinity tolerance. Annu. Rev. Plant Biol., 59: 651–681.
- Nazmy, A. and Arafa, F. (2017). Impact of amino acids complex foliar application on vegetative, growth, flowering traits and chemical constituents of marigold (*Calendula officinalis* L.) plants. World Journal of Agricultural Sciences, 13(6):247-256.
- Nofal, E.M.S.; El-Mahrouk, M.E.; El-Sayed, B.A. and Radwan, A.M.M. (2021). Effect of NPK fertilizer and some natural extract treatments on the chemical African composition ofmarigold (Tagetes L. var. dwarf erecta chrysanthemum). Applied Ecology and Environmental Research, 19(4):3153-3165.

- Nofal, F.H.; El-Sogai, M.U. and Seleem, A. (2015). Response of *Calendula officinalis* L. plants to growth stimulants under salinity stress. American J. Agric. and Environ. Sci., 15(9):1767-1778.
- Nosir, W.S. and El-Kinany, R.G. (2019). Effect of amino acids on growth and flowering of *Rosa hybrid* L. plant growing in nutrient film technique and solid substrate soilless culture. The Future Journal of Biology, 4:47-53.
- Osman, H.E. and Salem, O.M.A. (2011). Effect of seaweed extracts as foliar spray on sunflower yield and oil content. Egyptian J. of Phycol., 12:59-72.
- Phillips, I.D.J. (1971). Introduction to the Biochemistry and Physiology of Plant Grow Hormones. Mc. Growth- Hill Bock Company, USA, 173 p.
- Pramanick, B.; Brahmachari, K. and Gopsh, A. (2013). Effect of seaweed saps on growth and yield improvement of green gram. African J. of Agric. Res., 8:1180-1186.
- Prasad, A.; Chattopadhyay, A.; Chand, S.; Naqvi, A.A. and Yadav, A. (2006). Effect of soil sodicity on growth, yield, essential oil composition, and cation accumulation in rose-scented geranium. Communications in Soil Science and Plant Analysis, 37(13-14):1805-1817.
- Rafiee, H., Mehrafarin, A., Qaderi, A., Kalate, J.S. and Naghdi, B.H. (2013). Phytochemical, agronomical and morphological responses of pot marigold (*Calendula officinalis* L.) to foliar application of bio-stimulators (bioactive amino acid compounds). J. of Medicinal Plants, 12(47):48-61.
- Rahmatzadeh, S.; Khara, J. and Kazemitabar, S.K. (2012). Effects of tryptophan on growth and some physiological parameters in mycorrhiza inoculated plants of *Catharanthus roseus*, L. G. Don. International J. of Agric. Res. and Review, 2(5):564-572.

- Rehman, A.; Riaz, R.; Iqbal, M.; Shafi, J. and Ahmad, W. (2014). Responses of different morphological attributes of *Rosa hybrida* L. cv. Kardinal to saline water irrigation. Environmental Ecology and Research, 2(1):21-26.
- Roshani, S. and Asadi-Gharneh, H.A. (2019). Assessment effects of different level of amino acid and seaweed extract on growth traits and essence components of sweet scented geranium (*Pelargonium graveolens* L.). Journal of Crop Nutrition Science, 5 (2): 12-24.
- Russel, R.S. (1982). Plant Root Systems: Their Function and Interaction with the Soil. McGraw-Hill Education, USA, 298 p.
- Sahoo, D. (2000). Farming the ocean: Seaweeds Cultivation and Utilization. Aravali, New Delhi, India 44 p.
- Sewedan, E.; El–Naggar, H. and Osman, A. (2012). Effect of nitrogen and diphenylamine on *Gladiolus hybrid* cv. Sancerre production. J. of Hort. Sci. and Ornamental Plants, 4(3):267-274.
- Shahmoradi, H. and Naderi, D. (2018). Improving effects of salicylic acid on morphological, physiological and biochemical responses of salt-imposed winter jasmine. International Journal of Horticultural Science and Technology, 5(2):219-230.
- Shannon, M.C. and Grieve, C.M. (1999). Tolerance of vegetable crops to salinity, Scientia Horticulturae, 78:5-38.
- Shehata, S.M.; Abdel-Azem, H.S.; Abou El-Yazied, A. and El-Gizawy, A.M. (2011). Effect of foliar spraying with amino acids and seaweed extract on growth chemical constitutes, yield and its quality of celeriac plant. European Journal of Scientific Research, 58(2):257-265.
- Singh, K.N. and Chatrath, R. (2001). Salinity tolerance. In: Reynalds, M.P.; Ortiz-Monasterio, I. and McNab, A. (eds.), Application of Physiology in Wheat

- Breeding. CIMMYT, Mexico, pp. 101–110.
- Singh, P.; Pandey, S.S.; Dubey, B.K.; Raj, R.; Barnawal, D.; Chandran, A. and Ur Rahman, L. (2021). Salt and drought stress tolerance with increased biomass in transgenic *Pelargonium graveolens* through heterologous expression of ACC deaminase gene from *Achromobacter xylosoxidans*. Plant Cell, Tissue and Organ Culture, 147: 297–311.
- Sridhar, S. and Rengasamy, R. (2010). Effect of seaweed liquid fertilizer on the growth, biochemical constituents and yield of *Tagetes erecta*, under field trial. J. of Phytology, 2(6):61-68.
- Talib, M.; Hassan, A.E. and Hashim, A.O. (2012). Response of *Calendula officinalis* to spraying with algal extract. Al-Fourat J. Agric. Sci., 4(2):1-11.
- Tayama, H.K. and Carver, S.A. (1990). Zonal geranium growth and flowering responses to six growth regulators. HortScience, 25(1):82-83.
- Thirumaran, C.; Arumugam, M.; Arumugam, R. and Anantharaman, P. (2009a). Effect of seaweed liquid fertilizer on growth and pigment concentration of *Abelmoschus esculentus* (L) medikus. American-Eurasian Journal of Agronomy, 2(2):57-66.
- Thirumaran, C.; Arumugam, M.; Arumugam, R. and Anantharaman, P. (2009b). Effect of seaweed liquid pigment fertilizer on growth and concentration of Cyamopsis tetrogonolaba (L) Taub. American-Eurasian Journal of Agronomy, 2(2):57-
- Valdés, R.; Ochoa, J.; Franco, J.A.; Sánchez-Blanco, M.J. and Bañón, S. (2015). Saline irrigation scheduling for potted geranium based on soil electrical conductivity moisture and sensors. Agricultural Water Management, 149:123-130.

M.A.H. Abdou et al.

Walter, G.R. and Nawacki, E. (1987). Alkaloids Biology Metabolism in Plants. Planum Press., NY, USA, 152 p.

Wild, A. (1988). Russell's Soil Conditions and Plant Growth, 11th Ed. Longman Scientific and Technical, USA, 991 p.

Youssef, H.M.A. (1997). Physiological Studies on Some Annual Plants. M.Sc. Thesis, Fac. Agric., Moshtohor, Zagazig Univ, Egypt, 107 p.

تأثير بعض المواد المنشطة على نمو وتزهير نبات الجارونيا تحت ظروف ماء الري المالح

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تجارب أصبص تم إجرائها خلال موسمي نمو متعاقبين ٢٠١٩ و٢٠٢٠ بمشتل نباتات الزينة، قسم البساتين، كلية الزراعة، جامعة المنيا، مصر لدراسة استجابة نبات الجارونيا الوردي (Pelargonium x hortorum L.H. Bailey) النامي في التربة الرملية الجيرية لأربعة مستويات من الملوحة وثلاث معاملات للمنشطات الحيوية (مستخلص الأعشاب البحرية ، والأحماض الأمينية والكنترول) بالإضافة إلى التفاعل بينهما من حيث صفات النمو الخضري، وخصائص النمو الجذري، والصفات الزهرية وبعض المكونات الكيميائية. أشارت النتائج إلى أن صفات النمو الخضري (ارتفاع النبات، عدد الأفرع وعدد الأوراق/نبات)، معاملات نمو الجذور (وزن الجذر الطازج والجاف)، الصفات الزهرية (عدد النورات/نبات وقطر النورة)، وصبغات التمثيل الضوئي والنسبة المئوية لكل من النيتروجين والفوسفور والبوتاسيوم قد انخفضت معنوياً نتيجة للري بالمياه المالحة، وكانت أكبر الانخفاضات مع الريّ بماء مالح (٧٠,٠٠٪). في الوقت نفسه، أدت معاملات الملوحة إلى زيادة كل من النسبة المئوية للصوديوم والكالسيوم والكلوريد ومحتوى البرولين (مليجرام/جرام وزن طازج). بالإضافة إلَّى ذلك، أدى رش النباتات بأي من المنشطات الحيوية (مستخلص الأعشاب البحرية أو الأحماض الأمينية، بتركيز ٢٠٠ جزء في المليون لكل منهماً) إلى زيادة معنوية في جميع الصفات السابقة للنمو الخضري ونمو الجذور والصفات الزهرية وبعض المكونات الكيميائية (النسبة المئوية لكل من النيتروجين و الفوسفور و البوتاسيوم والكالسيوم) مقارنة بالكنترول. وبالتالي، قالت معاملات المنشطات الحيوية كل من النسبة المئوية لكل من الصوديوم والكلوريد وكذلك محتوى البرولين (مليجرام/جرام وزن طازج). تفوقت معاملات التداخل بين الري بماء الصنبور مع الحمض الأميني (٢٠٠ جزء في المليون) على معاملات التداخل الأخرى في كل الصفات المدروسة ماعدا النسبة المئوية للبو تاسيوم.