

EFFECT OF HUMIC ACID AND SOME MICRO NUTRIENTS ON GROWTH, SEED YIELD AND OIL CONTENT OF *BORAGO OFFICINALIS* L. PLANT

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ABSTRACT: Borage (*Borago officinalis* L.) is a yearly herbaceous plant belonging to the *Boraginaceae* family and known as the bee plant or bee bread. Borage is a significant medicinal plant indigenous to the Mediterranean region and has become widespread in many other countries. A factorial experiment was executed to assess the impact of humic acid and micro nutrients on the growth characteristics, chemical composition, and oil production of borage plants. It was carried out at the Experimental Farm of the Horticulture Department, Faculty of Agriculture, Moshtohor, Benha University, through the two consecutive growing seasons of 2020/2021 and 2021/2022. This trial comprised 16 treatments which were the outcome of the combination between soil addition with humic acid at 0 (control), 2, 3 and 4 kg/fed, and foliar spray with water (control), Zn, Fe or Mn at 100 ppm for each one. Results indicated that the treatment of humic acid at 4 kg/fed + Fe at 100 ppm was significantly improved the growth characters including number of branches, fresh and dry weight of herb. The treatment of humic acid at 4 kg/fed + Fe at 100 ppm also produced the greatest values of total nitrogen, phosphorus, potassium and carbohydrate in both seasons. Meanwhile, the treatment of humic acid at 4 kg/fed + Zn at 100 ppm significantly improved the characters including plant height, fresh and dry weights of inflorescences, and seed yield in both seasons. The treatment of humic acid at 4 kg/fed + Zn at 100 ppm gave the highest fixed oil percentage followed by humic acid at 4 kg/fed + Fe at 100 ppm. Exactly 68 compounds were identified and accounted in the components of *Borago officinalis* fixed-seed oil samples of the second season when analyzed by GC-MS. Where the sample resulted from the treatment of humic acid (4 kg/fed) + Fe (100 ppm) recorded 33 compounds. Conversely, the sample resulted from the control treatment recorded 14 compounds.

Keywords: Borage, humic acid, micro nutrients, fatty acid

INTRODUCTION

Borage (*Borago officinalis* L.) is an annual herbaceous plant belonging to the *Boraginaceae* family (Adamczyk-Szabela and Wolf, 2024). Also known as the bee plant or bee bread (El-Hafid *et al.*, 2002). The entire plant is covered with coarse, white, stiff bristles. Young leaves are edible or prepared like spinach, although this practice is

principally common for plants grown in domestic gardens. The flowers are vibrant both blue and shaped like stars, with conspicuous black anthers forming a cone. The fruit consists of 4 dark brown to black nutlets (Seifzadeh *et al.*, 2020). Borage is a medicinal plant indigenous to the Mediterranean region and has become widespread in many other countries (El-

Rahman *et al.*, 2023). Though native to Europe, Asia Minor, and North Africa, it is cultivated worldwide, with major producers being the UK, Canada, and New Zealand (Galambosi *et al.*, 2014). Its therapeutic properties are esteemed both in modern pharmaceuticals and traditional healing practices. It serves as a potent anti-inflammatory agent, aiding in avoiding colds, bronchitis, and respiratory ailments (Bulgari *et al.*, 2017 and Montaner *et al.*, 2022). Additionally, borage reduces amount of cholesterol and assists in controlling the gut and cardiovascular issues (Gupta and Singh 2010 & Sheikhzadeh *et al.*, 2021). It is used for medicinal and culinary purposes, with recent cultivation focusing on oil production (Asadi-Samani *et al.*, 2014). The agricultural regions for this plant are not clearly outlined. Varieties with blue flowers are prevalent, while those with white flowers are primarily used for culinary purposes (Galambosi *et al.*, 2014). The leaves and flowers comprise multiple bioactive combinations, comprising mucilage, tannin, saponins, pyrrolizidine alkaloids, V.C, Ca, and K (Gupta and Singh, 2010). Additionally, borage seeds yield gamma-linolenic acid, a highly valued dietary supplement (Gilbertson *et al.*, 2014). The global trade volume of borage seeds ranges from 1000 to 2000 tonnes annually, with considerable yearly fluctuations (Galambosi *et al.*, 2014).

Relying exclusively on synthetic fertilizers in continuous intensive cropping systems does not effectively sustain long-term crop productivity. However, incorporating organic amendments alongside chemical fertilizers can significantly enhance soil's physical characteristics, preserve higher soil fertility levels, and result in improved crop yields (Bera *et al.*, 2024). This strategy merges the quick nutrient supply offered by chemical fertilizers with the prolonged benefits of organic matter, such as better soil structure, increased water retention, and enhanced microbial activity (Elankavi *et al.*, 2020). Among organic compounds, humic acid is one of the most influential molecules, significantly impacting various agronomic

parameters and soil attributes (Ampong *et al.*, 2022). Humic acid is predominantly formed from the breakdown of plant and animal matter and plays several essential roles in agricultural systems. These roles include enhancing the biological and physical properties of soil qualities by getting better soil texture, structure, microbial activity, and moisture retention (Nardi *et al.*, 2021 and Shah *et al.*, 2018). Moreover, humic acid functions as a chelating agent, increasing the accessibility of trace components in the soil and facilitating greater nutrient absorption by plants. It also helps mitigate the uptake of hazardous heavy metals by plants (Wu *et al.*, 2017). The use of humic acid also boosts crop development by growing the levels of growth-regulating like cytokinin and auxin, which are essential for nutrient metabolism, stress adaptation, and photosynthetic performance (Canellas *et al.*, 2020 and Jindo *et al.*, 2020). Furthermore, humic acid positively influences plant cell membranes, improving mineral transport, enhancing protein synthesis, increasing enzyme activity, reducing the impact of harmful elements, and supporting a more robust microbial community (Khaled and Fawy, 2011).

Micro nutrients are vital trace elements that plants need in small amounts to achieve ideal development and reproduction. Nevertheless, these amounts are minimal, they are crucial for numerous biochemical and physiological functions. These elements play essential roles in processes such as enzyme activation, chlorophyll production, and nutrient assimilation, making them indispensable for the as a whole health and vigor of plants (Ahmed, 2024). Micronutrients are essential for sustaining the as a whole robustness and vigor of plants. They are integral to numerous physiological and biochemical functions, each contributing distinctively to the enhancement of plant health and efficiency (Aftab and Hakeem, 2020). These minor elements, usually present in low concentrations, are essential for the activation and support of enzymes that drive crucial metabolic pathways (Gomes *et al.*, 2020).

Iron (Fe) is not merely a fundamental element; it is crucial for a multitude of enzymatic processes. Its involvement is essential for the optimal functioning of biological mechanisms such as photosynthesis, cellular respiration, and nitrogen fixation (Li *et al.*, 2021). Decreased levels of iron can disrupt these systems, leading to reduced plant health (Herlihy *et al.*, 2020).

Similarly, zinc (Zn) is integral to various physiological processes, comprising nucleic acid synthesis and carbohydrate metabolism, making it indispensable for cellular activities (Baladrán-Valladares *et al.*, 2021). Zinc (Zn) is actively engaged in the biosynthesis of auxins and phytohormones that control cell expansion, root architecture, and the regulation of flowering initiation (Balafrej *et al.*, 2020; Otiende *et al.*, 2021 and Tripathi *et al.*, 2022). Disruptions in zinc concentrations can thus produce cascading effects on plant developmental processes and overall growth trajectories (Suganya *et al.*, 2020).

Manganese (Mn) is crucial for the biosynthesis of compounds that maintain cellular structural stability. Additionally, it serves as a critical component in reinforcing plant antioxidant mechanisms, thereby safeguarding against potential environmental stresses (Ghorbani *et al.*, 2019 and Ye *et al.*, 2019).

The main aim of this work is to study the effect of humic acid and micro nutrients on the growth characteristics, chemical composition, and oil content of borage plants (*Borago officinalis* L.).

MATERIALS AND METHODS

A factorial experiment was performed to assess the role of humic acid and micro nutrients on the growth characteristics, chemical composition, and oil production of borage plants. It was carried out at the Experimental Farm of the Horticulture Department, Faculty of Agriculture, Moshtohor, Benha University, during the two

consecutive growing seasons of 2020/2021 and 2021/2022.

Materials:

Borage seeds were directly sown in soil on the second of October 2020 and 2021 within plots measuring 1×1 meters. Each plot comprised 2 rows spaced 50 cm apart, with 50 cm between individual plants. Seeds of Borage were obtained from Horticulture Department Station, Faculty of Agriculture, Benha University. Fertilization involved the application of calcium superphosphate (15.5% P₂O₅) at a rate of 100 kg per feddan, ammonium sulfate (20.5% N) at 100 kg per feddan, and potassium sulfate (48% K₂O) at 50 kg per feddan (AzzEl-Din and Hendawy, 2010). Phosphorus was added during soil preparation, while nitrogen was applied in February and then in March, across both growing seasons. Potassium sulfate was added at three doses, the first one was added during soil preparation, the second one was added after two weeks from thinning, while the third dose was added after two weeks from the second one during two seasons.

The procedures outlined by Jackson *et al.* (1973) and Black *et al.* (1982) were utilized to assess the physical and chemical characteristics of the experimental soil. Table (1) summarizes the results of the soil analysis for both seasons. Furthermore, chemical analyses were conducted on the soil prior to the commencement of each season to evaluate factors such as organic matter content, calcium carbonate levels, available nitrogen, phosphorus, and potassium, as well as pH and electrical conductivity.

Layout of the experimental:

The experiment followed a randomized complete block design with three replications for each treatment, each treatment represented by three plots with four plants for each plot. This experiment included 16 treatments resulted from the combination between humic acid at addition (0, 2, 3 and 4 kg/fed, and spraying with water (control), Zn, Fe or Mn at 100 ppm for each one. Humic acid was added

Table 1. Chemical and physical characteristics of the experimental soil.

Parameters	2020/2021	2021/2022
Mechanical Properties		
Coarse sand (%)	8.9	5.9
Fine sand (%)	13.8	12.2
Loam (%)	22.1	23.0
Clay (%)	55.2	59.0
Textural class	Clay loam	Clay loam
Chemical Analysis		
Organic matter (%)	1.78	1.82
Calcium carbonate (%)	0.84	0.98
Available nitrogen (mg Kg ⁻¹)	0.67	0.73
Available phosphorus (mg kg ⁻¹)	0.43	0.39
Available potassium (mg kg ⁻¹)	198	209
pH water (1:1)	7.67	7.58
Electrical conductivity (dS/m)	0.95	0.75

as soil drench at three equal doses, the first one was done during soil preparation, the second one was conducted after one week from thinning, while the third one was carried out after two weeks from the second one during both seasons.

Three kinds of microelements, Zn EDTA (13%), Fe EDTA (13%) and Mn EDTA (13%) at 100 ppm were foliar sprayed early in the morning at three times, the first spray was carried out after one week from thinning and at three weeks intervals. Control plants were sprayed with distilled water.

Sampling:

Vegetative growth parameters were estimated just before flowering parameters were taken at full blooming in the two seasons. Besides, seed yield parameters were estimated at the end of the experiment. Chemical composition parameters were determined two weeks after the last spray treatments in the two seasons.

During each of the two growth seasons, the following traits were measured:

Growth, florescence and seed yield Parameters:

Plant height (cm), number of branches, vegetative fresh and dry weights (g/plant), inflorescences fresh and dry weights (g/plant), seed yield (kg/fed), fixed oil % and

fixed oil yield (l/fed) were determined. In order to prevent seed loss during maturation, the seeds were harvested as soon as they started to ripen, let to dry, and then weighed.

Chemical Composition:

Leaf N%, P%, K% and total carbohydrate were determined according to Horneck and Miller (1998), Hucker and Catroux (1980), Horneck and Hanson (1998) and Herbert *et al.* (1971), respectively.

Extraction of fat, and determination of fatty acids of oil:

The AOAC (1984) techniques were followed in order to extract a fixed oil percentage from borage seed using hexane in a Soxhlet system HT apparatus.

Gas chromatography-mass spectrometry (GC/MS) analysis:

A GC (Agilent Technologies 7890A) with a mass-selective detector running on an HP-5ms capillary column (30 µm x 0.25 mm i.d. and 0.25 µm film thickness) was used at the Regional Center for Food and Feed (RCFF), ARC, Giza, Egypt to determine the methanolic extract. At a pace of 3 degrees Celsius per minute, the temperature was raised from 80 to 230 degrees. Helium was the carrier gas, flowing at a rate of 1 milliliter per minute. The process of bioactive chemical identification involved computer matching

with the National Institute Standard and Technique database, as well as comparing the mass spectra and retention times of the compounds with those of genuine standards.

Statistical analysis:

Analysis of variance (ANOVA) was used to examine the values of all the data that were gathered as part of factorial investigations carried out in a complete randomized block design. LSD_{5%} test and Duncan multiple range test were used to differentiate means according to Snedecor and Cochran (1991).

RESULTS AND DISCUSSION

It is clear that treating *Borago officinalis* plants with humic acid and some micro nutrients increased vegetative growth, flowering, seed yield and chemical composition parameters compared with the control (without any addition).

Vegetative Characteristics:

Table (2) findings clearly show that the plant height was greatly affected by the utilization of humic acid and some micro nutrients. Results suggested that the plant height of borage was significantly ($P < 0.05$) affected by zinc or Manganese or Iron in addition to humic acid at different levels. It was found that studied fertilization treatments progressively heightened the plant height of *Borago officinalis* L. plant when compared with control in both seasons. Nevertheless, the treatment of humic acid at 4 kg/fed + Zn at 100 ppm had the positive effect. These results agree with those obtained by Memon *et al.* (2014) on *Phlox Paniculata* plants, Mohamed and Ghatas (2020) on *Salvia hispanica*, Omar (2020) on caraway plants, Vafa *et al.* (2020) on summer savory plants, Zghair *et al.* (2021) on *Rosmarinus officinalis* plants, Tawfik (2022) on *Foeniculum vulgare*, Hoseini *et al.* (2023) on *Ocimum basilicum*, Kazemi *et al.* (2023) on *Physalis alkekengi* plant, Khosravi *et al.* (2023) on *Salvia officinalis* plants, Korani *et al.* (2023) on *Cichorium intybus*, Mubarak *et al.* (2023) on Japanese cabbage Plants.

The treatment of humic acid at 4 kg/fed + Fe at 100 ppm significantly improved the growth characters including number of branches, fresh and dry weights of herb. In this respect, the greatest number of branches were 23.04 and 24.29 in the first and second seasons respectively, when compared with control (12.92 and 13.12). The highest increment in fresh weight of herb was 1102 and 1410 g/plant, respectively in both seasons when compared with control (713.33 and 885 g/plant, respectively). As well as that dry weight of herb was 197.33 and 266 g/plant, respectively in both seasons when compared with untreated plants (control) (127 and 164.3 g/plant).

These results agree with those obtained by El-Gohary *et al.* (2014) on *Mentha piperita*, Azizi and Safaei (2017) on *Nigella sativa*, Dukpa *et al.* (2017) on *Ipomoea reptans*, Aghdasi *et al.* (2018) on *Vigna radiata*, Bastani *et al.* (2018) on tobacco plants, Ayobizadeh *et al.* (2019) on sesame cultivars, Cieschi *et al.* (2019) on *Glycine max*, Davoodi *et al.* (2020) on *Nigella sativa*, Simsek and Çelik (2021) on *Spinacia oleracea*, Hayati *et al.* (2022) on *Nigella sativa*, Turan *et al.* (2022) on *Spinacia oleracea*.

Chemical composition:

Total nitrogen percentage:

Data displayed in Table (3) suggested that, the total nitrogen percentage was profoundly impacted by the humic acid and some micro nutrients treatments; it was found that studied fertilization treatments progressively increased the total nitrogen percentage of *Borago officinalis* L. plant when compared with control in both seasons of study. However, the treatment of humic acid at 4 kg/fed + Fe at 100 ppm produced the maximum total nitrogen percentage in both seasons (2.727, 2.773) respectively, subsequently in descending order by utilizing the treatment of humic acid at 4 kg/fed + Zn at 100 ppm (2.693, 2.680), respectively in the

Table 2. Effect of humic acid and some micro nutrients on vegetative growth of *Borago officinalis* plants during 2020/2021 and 2021/2022 seasons.

Humic acid treatments	Micronutrient treatments									
	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean
	Plant Height (cm)					No. of branches/ plant				
	First season									
0 (Control)	81.79 ^I	84.27 ^{FG}	83.72 ^{GH}	82.91 ^H	83.17 ^D	12.92 ^I	13.75 ^I	15.52 ^{FG}	14.91 ^{GH}	14.28 ^D
2 kg/fed	84.41 ^{FG}	88.05 ^E	85.24 ^F	84.98 ^F	85.67 ^C	14.64 ^H	16.28 ^F	17.79 ^E	16.33 ^F	16.26 ^C
3 kg/fed	90.61 ^D	97.04 ^B	94.05 ^C	93.04 ^C	93.69 ^B	16.21 ^F	18.04 ^E	22.01 ^B	20.56 ^C	19.21 ^B
4 kg/fed	93.24 ^C	104.85 ^A	96.04 ^B	94.03 ^C	97.04 ^A	17.58 ^E	19.24 ^D	23.04 ^A	21.84 ^B	20.43 ^A
Mean	87.51 ^D	93.55 ^A	89.76 ^B	88.74 ^C		15.34 ^D	16.83 ^C	19.59 ^A	18.41 ^B	
LSD_{0.05}	A: 0.5356	B: 0.6634	A×B: 1.071			A: 0.4313	B: 0.5471	A×B: 0.8675		
	Second season									
0 (Control)	85.36 ^J	92.30 ^{FG}	89.96 ^{HI}	89.03 ^I	89.16 ^D	13.12 ^K	15.02 ^J	17.09 ^{GH}	16.14 ^{HI}	15.34 ^D
2 kg/fed	91.08 ^{GH}	97.02 ^{DE}	93.75 ^F	90.88 ^{GH}	93.18 ^C	15.65 ^{IJ}	15.87 ^{IJ}	19.06 ^{CDE}	18.05 ^{EFG}	17.16 ^C
3 kg/fed	96.21 ^E	106.70 ^B	103.22 ^C	97.98 ^D	101.03 ^B	17.18 ^{FG}	18.28 ^{DE}	23.43 ^A	19.66 ^C	19.64 ^B
4 kg/fed	103.47 ^C	112.09 ^A	108.21 ^B	104.63 ^C	107.10 ^A	18.14 ^{EF}	19.31 ^{CD}	24.29 ^A	20.85 ^B	20.65 ^A
Mean	94.03 ^D	102.03 ^A	98.79 ^B	95.63 ^C		16.02 ^D	17.12 ^C	20.97 ^A	18.68 ^B	
LSD_{0.05}	A: 0.2664	B: 0.8726	A×B: 1.686			A: 0.5173	B: 0.5977	A×B: 1.035		
	Fresh weight of herb/plant (g)					Dry weight of herb/ plant (g)				
	First season									
0 (Control)	713.33 ^M	801.67 ^I	792.67 ^J	763.67 ^L	767.84 ^D	127.00 ^L	145.00 ^I	141.67 ^J	136.00 ^K	137.42 ^D
2 kg/fed	782.00 ^K	892.67 ^F	898.33 ^F	817.67 ^H	847.67 ^C	140.67 ^J	161.00 ^F	161.33 ^F	146.00 ^I	152.25 ^C
3 kg/fed	871.00 ^G	983.33 ^C	986.33 ^C	937.00 ^E	944.42 ^B	155.00 ^H	174.67 ^D	175.00 ^D	167.00 ^E	167.92 ^B
4 kg/fed	892.00 ^F	1072.67 ^B	1102.00 ^A	963.00 ^D	1007.42 ^A	158.33 ^G	195.00 ^B	197.33 ^A	184.67 ^C	183.83 ^A
Mean	814.58 ^D	937.59 ^B	944.83 ^A	870.34 ^C		145.25 ^C	168.92 ^A	168.83 ^A	158.42 ^B	
LSD_{0.05}	A: 3.535	B: 3.957	A×B: 7.070			A: 1.130	B: 1.698	A×B: 2.260		
	Second season									
0 (Control)	885 ^O	963 ^K	943 ^L	914.67 ^N	926.42 ^D	164.3 ^I	181.7 ^G	186.3 ^F	173.7 ^H	176.5 ^D
2 kg/fed	924.33 ^M	1109.33 ^H	1083 ^I	984.33 ^J	1025.25 ^C	175.7 ^H	209.7 ^D	204 ^E	185.3 ^{FG}	193.68 ^C
3 kg/fed	1080.33 ^I	1311 ^C	1293.67 ^D	1117.67 ^G	1200.67 ^B	204.3 ^E	247.3 ^B	246 ^B	211.7 ^D	227.33 ^B
4 kg/fed	1124.33 ^F	1383.67 ^B	1410 ^A	1217 ^E	1283.75 ^A	211.7 ^D	262.3 ^A	266 ^A	233.3 ^C	243.33 ^A
Mean	1003.50 ^D	1191.75 ^A	1182.42 ^B	1058.42 ^C		189 ^C	225.25 ^A	225.58 ^A	201 ^B	
LSD_{0.05}	A: 2.887	B: 6.158	A×B: 5.773			A: 1.877	B: 2.622	A×B: 3.755		

Table 3. Effect of humic acid and some micro nutrients on chemical composition of *Borago officinalis* plants during 2020/2021 and 2021/2022 seasons.

Humic acid treatments	Micro nutrient treatments									
	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean
	N %					P %				
	First season									
0 (Control)	2.143 ^N	2.210 ^L	2.237 ^K	2.180 ^M	2.193 ^D	0.222 ^H	0.219 ^H	0.227 ^{GH}	0.218 ^H	0.222 ^D
2 kg/fed	2.327 ^J	2.377 ^I	2.417 ^H	2.497 ^G	2.405 ^C	0.232 ^{FGH}	0.247 ^{CDEF}	0.241 ^{DEFG}	0.239 ^{EFG}	0.240 ^C
3 kg/fed	2.571 ^F	2.617 ^C	2.587 ^{EF}	2.593 ^{DE}	2.592 ^B	0.252 ^{BCDE}	0.259 ^{ABC}	0.263 ^{ABC}	0.258 ^{ABCD}	0.258 ^B
4 kg/fed	2.617 ^C	2.693 ^B	2.727 ^A	2.610 ^{CD}	2.662 ^A	0.265 ^{AB}	0.269 ^{AB}	0.272 ^A	0.267 ^{AB}	0.268 ^A
Mean	2.415 ^C	2.474 ^B	2.492 ^A	2.470 ^B		0.2428 ^A	0.2485 ^A	0.2508 ^A	0.2457 ^A	
LSD_{0.05}	A: 0.008426		B: 0.009989		A×B: 0.01685		A: 0.008426		B: 0.009989	
	Second season									
0 (Control)	2.197 ^L	2.283 ^H	2.257 ^I	2.237 ^{JK}	2.244 ^D	0.208 ^F	0.212 ^F	0.220 ^{EF}	0.213 ^F	0.213 ^C
2 kg/fed	2.250 ^J	2.230 ^K	2.343 ^G	2.290 ^H	2.278 ^C	0.219 ^{EF}	0.231 ^{DE}	0.239 ^{CD}	0.219 ^{EF}	0.227 ^B
3 kg/fed	2.483 ^F	2.567 ^E	2.627 ^C	2.597 ^D	2.569 ^B	0.243 ^{BCD}	0.242 ^{BCD}	0.257 ^{AB}	0.253 ^{ABC}	0.248 ^A
4 kg/fed	2.587 ^D	2.680 ^B	2.773 ^A	2.563 ^E	2.651 ^A	0.250 ^{BC}	0.257 ^{AB}	0.268 ^A	0.250 ^{BC}	0.256 ^A
Mean	2.379 ^D	2.440 ^B	2.500 ^A	2.422 ^C		0.230 ^B	0.236 ^B	0.246 ^A	0.234 ^B	
LSD_{0.05}	A: 0.008426		B: 0.009989		A×B: 0.01685		A: 0.008426		B: 0.009989	
	K %					Total Carbohydrate %				
	First season									
0 (Control)	1.340 ^K	1.387 ^J	1.410 ^I	1.347 ^K	1.371 ^D	14.87 ^L	15.27 ^K	15.77 ^J	14.93 ^L	15.21 ^D
2 kg/fed	1.390 ^J	1.487 ^G	1.587 ^D	1.463 ^H	1.482 ^C	15.90 ^L	16.87 ^H	17.07 ^G	16.37 ^I	16.55 ^C
3 kg/fed	1.463 ^H	1.517 ^F	1.670 ^B	1.533 ^{EF}	1.546 ^B	17.17 ^{FG}	17.93 ^D	18.13 ^C	17.27 ^F	17.63 ^B
4 kg/fed	1.527 ^{EF}	1.637 ^C	1.720 ^A	1.543 ^E	1.607 ^A	17.67 ^E	18.40 ^B	18.77 ^A	18.10 ^C	18.24 ^A
Mean	1.430 ^D	1.507 ^B	1.597 ^A	1.472 ^C		16.40 ^D	17.12 ^B	17.44 ^A	16.67 ^C	
LSD_{0.05}	A: 0.008426		B: 0.009989		A×B: 0.01685		A: 0.07050		B: 0.1094	
	Second season									
0 (Control)	1.513 ^{DE}	1.463 ^{EF}	1.480 ^{EF}	1.433 ^F	1.472 ^C	15.33 ^L	15.97 ^J	16.33 ^{HI}	15.63 ^K	15.82 ^D
2 kg/fed	1.463 ^{EF}	1.513 ^{DE}	1.500 ^{EF}	1.470 ^{EF}	1.487 ^C	16.23 ^I	16.77 ^G	17.27 ^F	16.43 ^H	16.68 ^C
3 kg/fed	1.523 ^{CDE}	1.580 ^{BCD}	1.623 ^{AB}	1.597 ^{BC}	1.581 ^B	17.80 ^E	18.27 ^D	18.93 ^B	18.10 ^D	18.28 ^B
4 kg/fed	1.590 ^{BC}	1.633 ^{AB}	1.687 ^A	1.623 ^{AB}	1.633 ^A	18.10 ^D	18.27 ^D	19.20 ^A	18.47 ^C	18.51 ^A
Mean	1.522 ^B	1.547 ^{AB}	1.573 ^A	1.531 ^B		16.87 ^D	17.32 ^B	17.93 ^A	17.16 ^C	
LSD_{0.05}	A: 0.03768		B: 0.04467		A×B: 0.07536		A: 0.08426		B: 0.08358	

both seasons, when compared with control. The lowest value of this parameter was acquired by the treatment of control (2.143, 2.197) respectively, in both seasons.

Total phosphorus percentage:

Data displayed in Table (3) demonstrated that, the total phosphorus percentage was positively affected by the humic acid and some micro nutrients treatments, it was found that studied fertilization treatments progressively increased the total phosphorus percentage of *Borago officinalis* L. plant when compared with control in both seasons. The treatment of humic acid at 4 kg/fed + Fe at 100 ppm produced the maximum total phosphorus in both seasons (0.272, 0.268), respectively, descending order by using the treatment of humic acid at 4 kg/fed + Zn at 100 ppm (0.269, 0.257), respectively at the first and second seasons, when compared with control. The least values of this parameter were listed by control (0.222 and 0.208), respectively, in both seasons.

Total potassium percentage:

Data presented in Table (3) indicated that, the total potassium percentage was greatly affected by the humic acid and some micro nutrients treatments; it was found that studied fertilization treatments progressively increased the total potassium percentage of *Borago officinalis* L. plant when compared with control in both seasons of study. However, the treatment of humic acid at 4 kg/fed + Fe at 100 ppm produced the maximum total potassium in both seasons (1.720, 1.687), respectively, followed in descending order by using the treatment of humic acid at 4 kg/fed + Zn at 100 ppm (1.637, 1.633), respectively at the first and second seasons, when compared with control. The lowest value of this parameter was gained by using the treatment of control (1.340, 1.513), respectively, in both seasons.

Total Carbohydrate percentage:

Data offered in Table (3) demonstrated that, the total carbohydrate percentage was enormously affected by the humic acid and some micro nutrient treatments; it was found

that studied fertilization treatments progressively increased the total carbohydrate percentage of *Borago officinalis* L. plant when compared with control in both seasons. The treatment of humic acid at 4 kg/fed + Fe at 100 ppm produced the maximum total carbohydrate in both seasons (18.77 and 19.20, respectively), subsequently in a descending sequence by utilizing the treatment of humic acid at 4 kg/fed + Zn at 100 ppm (18.40 and 18.27, respectively in the first and second seasons), when compared with control. The lowest value of this parameter was gained by using the treatment of control (14.87 and 15.33, respectively, in both seasons).

Flowering growth and seeds yield:

From the results listed in Table (4) it is evident that the treatment of humic acid at 4 kg/fed + Zn at 100 ppm significantly improved the characteristics including fresh as well as dry weight of inflorescences, and seed yield in both seasons. The highest increment in fresh weight of inflorescences was 247 and 252 g/plant, respectively in both seasons when compared with control (183.33 and 232 g/plant, respectively). As well as, dry weight of inflorescences was 37.57 and 40.27 g/plant, respectively in both seasons when compared with control (27.20 and 28.93 g/plant). Additionally, in both seasons, it generally showed a greater and statistically significant increase in seed output per faddan when compared to the other ones under examination. It produced 378 and 387 kg/fed respectively, when compared with untreated plants (control) (286 and 369 kg/fed in the first and second seasons, respectively).

Fixed oil percentage and Fixed oil yield/fed (l):

According to the data in Table (5), utilizing humic acid and some micro nutrients treatments had a more significant effect on the fixed oil percentage of borage seeds than the control in both seasons. The treatment of humic acid at 4 kg/fed + Zn at 100 ppm gave the highest fixed oil percentage at 28.57 and 26.90% followed (in descending order) by

Table 4. Effect of humic acid and some micro nutrients on flowering growth and seeds yield of *Borago officinalis* plants during 2020/2021 and 2021/2022 seasons.

Humic acid treatments	Micro nutrient treatments																	
	Fresh W. of flower/plant (g)						Dry W. of flower/plant (g)						Seed yield/ fed (kg)					
	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean			
0 (Control)	183.3 ^L	196.0 ^{HI}	191.3 ^{JK}	189.0 ^K	189.9 ^D	27.2 ^J	29.6 ^G	28.4 ^{HI}	28.1 ^I	28.3 ^D	286 ^H	297 ^{FG}	293 ^{GH}	290 ^{GH}	292 ^C			
2 kg/fed	193.3 ^{JU}	206.7 ^G	197.0 ^H	194.0 ^{IJ}	197.7 ^C	28.1 ^I	31.4 ^F	29.4 ^G	28.8 ^H	29.4 ^C	292 ^{GH}	321 ^E	318 ^E	304 ^F	309 ^B			
3 kg/fed	213.0 ^E	233.0 ^B	212.3 ^{EF}	209.7 ^F	217.0 ^B	31.4 ^F	34.8 ^B	31.4 ^F	31.7 ^F	32.3 ^B	356 ^{CD}	364 ^B	359 ^{BCD}	352 ^D	358 ^A			
4 kg/fed	217.0 ^D	247.0 ^A	228.0 ^C	217.3 ^D	227.3 ^A	32.9 ^D	37.6 ^A	33.9 ^C	32.4 ^E	34.2 ^A	352 ^D	378 ^A	364 ^B	361 ^{BC}	364 ^A			
Mean	201.67 ^C	220.67 ^A	207.17 ^B	202.5 ^C	202.5 ^C	29.9 ^D	33.3 ^A	30.8 ^B	30.2 ^C	30.2 ^C	322 ^D	340 ^A	333.5 ^B	326.75 ^C				
LSD_{0.05}	A: 1.4	B: 1.9	A×B: 2.9	A: 0.3	B: 0.3	A×B: 0.5	A: 3.8	B: 6.5	A×B: 7.7									
0 (Control)	185.3 ^K	198.7 ^I	196.7 ^I	192.7 ^J	193.3 ^D	28.9 ^J	31.6 ^H	31.2 ^{HI}	30.5 ^I	30.6 ^D	294 ^K	312 ^H	308 ^I	302 ^J	304 ^D			
2 kg/fed	195.7 ^{JU}	216.0 ^G	217.7 ^G	207.3 ^H	209.2 ^C	31.4 ^H	34.3 ^F	34.6 ^F	33.2 ^G	33.4 ^C	326 ^G	341 ^E	337 ^F	326 ^G	333 ^C			
3 kg/fed	224.0 ^F	237.7 ^C	231.7 ^D	228.3 ^E	230.4 ^B	35.7 ^E	38.3 ^C	36.5 ^{DE}	36.3 ^{DE}	36.7 ^B	358 ^D	372 ^{BC}	369 ^C	361 ^D	365 ^B			
4 kg/fed	232.0 ^D	252.0 ^A	245.7 ^B	237.0 ^C	241.7 ^A	37.0 ^D	40.3 ^A	39.3 ^B	38.6 ^{BC}	38.8 ^A	369 ^C	387 ^A	375 ^B	371 ^C	376 ^A			
Mean	209.2 ^D	226.1 ^A	222.9 ^B	216.3 ^C	216.3 ^C	33.3 ^D	36.1 ^A	35.4 ^B	34.7 ^C	34.7 ^C	337 ^D	353 ^A	347 ^B	340 ^C				
LSD_{0.05}	A: 1.5	B: 2.7	A×B: 3.0	A: 0.4	B: 0.4	A×B: 0.8	A: 1.9	B: 3.4	A×B: 3.8									

Table 5. Effect of humic acid and some micro nutrients on fixed oil % and fixed oil yield/fed (l) of *Borago officinalis* plants during 2020/2021 and 2021/2022 seasons.

Humic acid treatments	Micro nutrient treatments											
	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean	0 (Control)	Zn 100 ppm	Fe 100 ppm	Mn 100 ppm	Mean		
	Fixed oil %					Fixed oil yield/fed (l)						
	First season											
0 (Control)	24.60 ^K	25.80 ^{HI}	25.93 ^{GH}	25.63 ^I	25.49 ^D	70.43 ^P	76.60 ^L	75.93 ^M	74.07 ^N	74.26 ^D		
2 kg/fed	25.17 ^J	26.30 ^E	26.10 ^{FG}	25.97 ^{GH}	25.89 ^C	73.33 ^O	84.40 ^I	83.07 ^J	78.73 ^K	79.88 ^C		
3 kg/fed	25.97 ^{GH}	27.80 ^C	27.33 ^D	27.47 ^D	27.14 ^B	89.67 ^H	101.3 ^C	98.03 ^E	96.43 ^F	96.36 ^B		
4 kg/fed	26.23 ^{EF}	28.57 ^A	28.13 ^B	27.93 ^C	27.72 ^A	92.23 ^G	107.8 ^A	102.4 ^B	100.8 ^D	100.81 ^A		
Mean	25.49 ^D	27.12 ^A	26.87 ^B	26.75 ^C		81.42 ^D	92.53 ^A	89.85 ^B	87.51 ^C			
LSD_{0.05}	A: 0.0843		B: 0.1548		A×B: 0.1685		A: 0.1032		B: 0.07064		A×B: 0.2064	
	Second season											
0 (Control)	23.97 ^K	24.97 ^H	24.67 ^I	24.47 ^J	24.52 ^D	70.40 ^O	77.73 ^L	75.87 ^M	73.77 ^N	74.44 ^D		
2 kg/fed	24.67 ^I	25.33 ^F	25.17 ^G	25.30 ^{FG}	25.12 ^C	80.13 ^K	86.53 ^H	84.70 ^I	83.37 ^J	83.68 ^C		
3 kg/fed	25.30 ^{FG}	26.43 ^C	26.50 ^C	26.27 ^D	26.13 ^B	90.70 ^G	98.33 ^C	97.90 ^D	94.70 ^F	95.41 ^B		
4 kg/fed	25.77 ^E	26.90 ^A	26.73 ^B	26.50 ^C	26.48 ^A	95.37 ^E	103.4 ^A	100.2 ^B	98.07 ^D	99.26 ^A		
Mean	24.93 ^D	25.91 ^A	25.77 ^B	25.63 ^C		84.15 ^D	91.50 ^A	89.67 ^B	87.48 ^C			
LSD_{0.05}	A: 0.0705		B: 0.1094		A×B: 0.1410		A: 0.0923		B: 0.1139		A×B: 0.1846	

humic acid at 4 kg/fed + Fe at 100 ppm at 28.13 and 26.73%. On the other hand, the control is the lowest levels of this factor during both seasons (24.60 and 23.97%).

Also, the maximum and significant increases in fixed oil yield/fed (l) were obtained by the treatment of humic acid at 4 kg/fed + Zn at 100 ppm which gave the highest yield at 107.8 and 103.4 l followed (in descending order) by humic acid at 4 kg/fed + Fe at 100 ppm at 102.4 and 100.2 l. On the other hand, the control yielded the lowest levels of this factor during both seasons (70.43 and 70.40 l) during the first and second seasons.

Main Components of the *Borago Officinalis* L. seed-oil as analyzed and identified by GC-MS:

The main components of *Borago Officinalis* fixed-seed oil (as identified by GC-MS) when affected by humic acid (4 kg/fed) as soil application and foliar spray with Zn, Fe, or Mn at 100 ppm concentration in the second season (2021/2022) are

presented in Table (6) and Figs. (1, 2, 3, 4 and 5). While admitting that the result data in Table (6) were not subjected to statistical analysis, still there were clear indicators in the general trends of the obtained results which will be highlighted and stressed in the paragraphs. Exactly 68 compounds were identified and accounted in the components of *Borago Officinalis* fixed-seed oil samples of the second season when analyzed by GC-MS. For the fixed-seed oil resulting from treatment of humic acid (4 kg/fed) + Zn (100 ppm), the number of main compounds identified in plenty were 21 compounds, i.e., 11,13-Eicosadienoic acid methyl ester (2.95%), 1-Decanol (4.63%), 1-Nonyne (1.86 %), 1-Octadecyne (1.16 %), 2-Undecenal (2.65 %), 6,9,12-Octadecatrienoic acid, methyl ester (1.02 %), 7,10-Octadecadienoic acid, methyl ester (41.1%), 8,11,14-Docosatrienoic acid, methyl ester (0.83%), 9,12-Octadecadienoic acid (1.13%), 9-Octadecyne (0.8 %), Camphor (0.89%), cis-11-Eicosenoic acid (7.39 %), Dodecane (1.48 %), Erucic acid

Table 6. Main Components of the *Borago Officinalis* L. seed-oil as analyzed and identified by GC-MS when affected by Humic acid (4 kg/fed) as soil application and foliar spray with Zn, Fe, or Mn at 100 ppm concentration.

Components	Control		Humic acid (4 kg/fed) + Zn (100 ppm)		Humic acid (4 kg/fed) + Fe (100 ppm)		Humic acid (4 kg/fed) + Mn (100 ppm)		Humic acid (4 kg/fed)	
	RT (min)	Area sum %	RT (min)	Area sum %	RT (min)	Area sum %	RT (min)	Area sum %	RT (min)	Area sum %
11,13-Eicosadienoic acid, methyl ester	17.752	3.81	17.7	2.95	3.3	2.5			17.768	2.19
1-Decanol	5.847	3.14	5.7	4.63	5.7	6.23	5.7	1.1	5.712	4.55
1-Decyne	9.275	0.5			9.2	1.94				
1-Docosanol							14.4	1.65		
1-Dodecene					8.9	0.74				
1-Eicosanol							7.7	1.25		
1-Hexadecanol, 2-methyl-									9.2	1.52
1-Nonyne			7.7	1.86						
1-Octadecyne			11	1.16	10.8	2.03				
1-Octanol					3.5	4.37				
1-Tetradecanol					9	0.81	7.2	0.98		
1-Undecen-10-al					11	2.2				
2(10)-Pinene					13.8	6.17	13.8	2.12		
2-Decenal, (E)-										
2-Dodecenal, (E)-					11.1	0.97			7.1	2.12
2-Myristyonic acid							10.8	0.79		
2-Undecenal	9.821	1.84	9.8	2.65	9.8	3.65	9.8	1.22	9.821	1.49
2-Undecenal, E-									11	1.32
3,5-Octadiene, 4,5-diethyl-					11.4	3.01				
3-Pinanol					7	0.84				
6,8-Dimethoxy-4-methyl-4H-					12.7	2.32	12.6	1.56		
6,9,12-Octadecatrienoic acid, methyl ester			17.6	1.02						
7,10-Octadecadienoic acid, methyl ester	18.359	49.07	18.8	41.1			18.65	35.66	18.576	42.41
7-Hexadecyne					9.5	0.87	9.5	1.82		
7-Tetradecene					7.1	3.11				
8,11,14-Docosatrienoic acid, methyl ester	19.458	1.62	19.4	0.83			19.3	1.18	19.376	2.65
9,12-Octadecadienoic acid(Z,Z)-			12.3	1.13						
9-Octadecenoic acid (Z)-							15.5	0.94		
9-Octadecenoic acid (Z)-,							4.4	2.38		
9-Octadecyne	10.214	0.62	10.2	0.8	10.2	8.66	10.2	1.22	10.218	0.63
Benzyl oxy tridecanoic acid			3.4	1.72						
Camphor	7.811	1.21	10.4	0.89	10.5	7.11	10.5	1.91	10.489	1.38
cis-11-Eicosenoic acid	20.532	3.91	20.5	7.39			20.6	6.44	20.586	4.9
cis-7-Hexadecenoic acid							9	0.94		
Cyclododecene					9.4	1.35				
Cyclohexane, 1,1,3-trimethyl-					3.9	2.7				
Dodecane			7.1	1.48						
Erucic acid	23.12	1.47	23	1.65			23	2.55	23.047	2.13
Heneicosane					12.4	1.51				
Heptanoic acid					6.2	2.61				
Hexadecanoic acid, methyl ester							15.2	0.49		
Isopropyl linoleate							17.2	2.64		
Methoprene									21.4	1.22
Methyl γ-linolenate			13.2	1.87	13.3	5.28			13.302	0.71
Myristic acid							15.1	0.66		
Myristic acid, methyl ester	16.353	0.67	16.3	1.11					16.362	1.43

Continued

Table 6. Continued.

Components	Control		Humic acid (4 kg/fed) + Zn (100 ppm)		Humic acid (4 kg/fed) + Fe (100 ppm)		Humic acid (4 kg/fed) + Mn (100 ppm)		Humic acid (4 kg/fed)	
	RT (min)	Area sum %	RT (min)	Area sum %	RT (min)	Area sum %	RT (min)	Area sum %	RT (min)	Area sum %
Myristoleic acid					7.6	0.8				
Myrtenoic acid, butyl ester					14.5	4.89				
Neopentane					8.5	1.63				
n-Hexadecanoic acid	16.792	21.12	16.9	16.79			17	10.58	16.895	17.53
Nonane					4.8	1.02			3.5	1.5
Octanoic acid	8.648	2.41			8.7	1.63				
Oleic Acid							11.7	0.52	17.3	0.65
Oxiraneoctanoic acid, 3-6ctyl-, cis-							12.3	0.91		
o-Xylene					4.4	4.33			4.4	1.73
Palmitic acid, 2-(tetradecyloxy)ethyl ester									15.8	1.07
Palmitoleic acid							11.9	0.46		
Pentadecanoic acid			15.8	1.57			16.2	13.56		
Phytol							12.1	2.15		
Pinolenic acid					15.7	9.85				
p-Xylene					4.3	1.65				
Rescinamine									15.4	0.47
Retinal									12.1	0.52
Tetradecanoic acid	14.939	4.98	14.9	1.8					14.967	2.63
Tridecanedial							13.3	0.85		
α-Humulene nonanal			13.6	5.62					13.643	3.25
1-Decene					7.7	1.65	6.8	1.46		
					5.5	1.57				

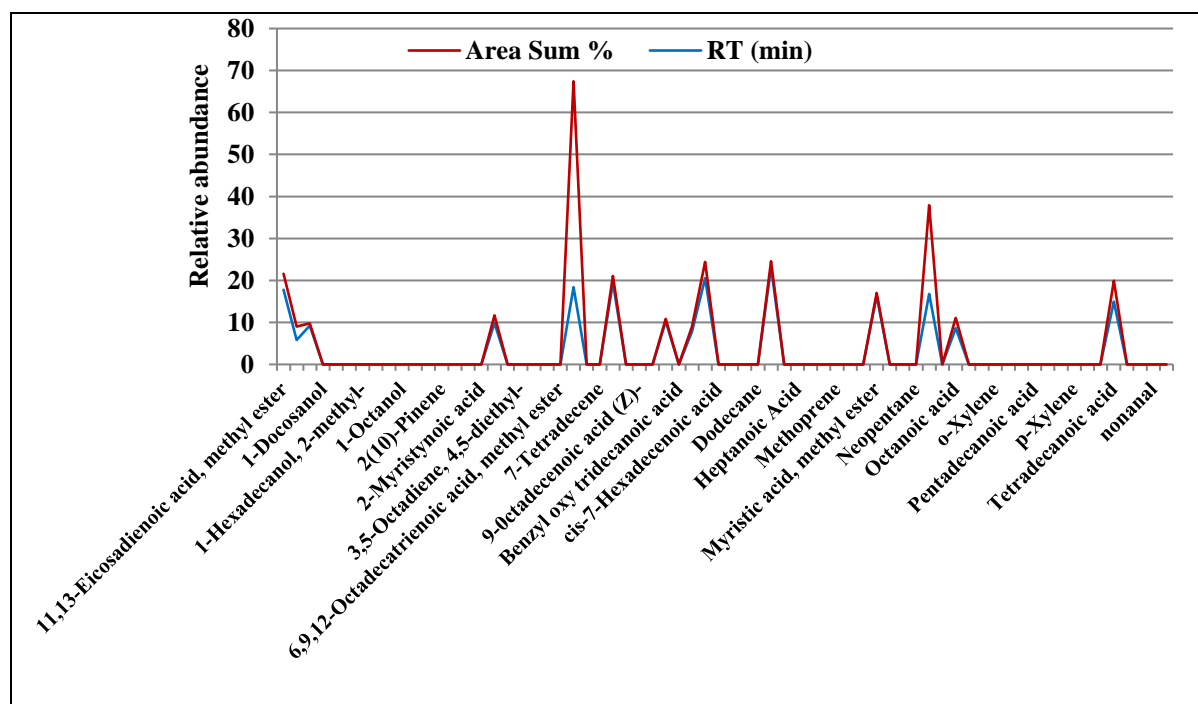


Fig. 1. Chart of GC-mass chromatogram of *Borago Officinalis* L. seed-oil from the plants treated with only water (Control) during the second season (2021/2022).

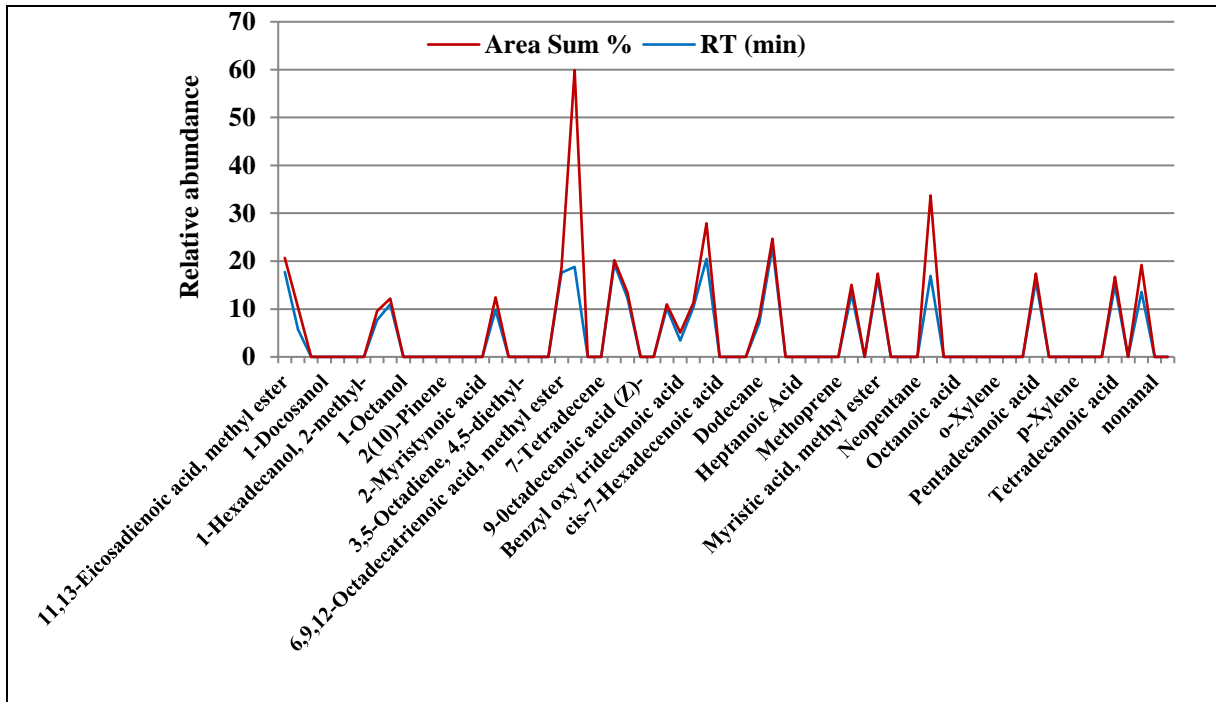


Fig. 2. Chart of GC-mass chromatogram of *Borago Officinalis* L. seed oil from the plants treated with humic acid (4 kg/fed) + Zn (100 ppm) during the second season (2021/2022).

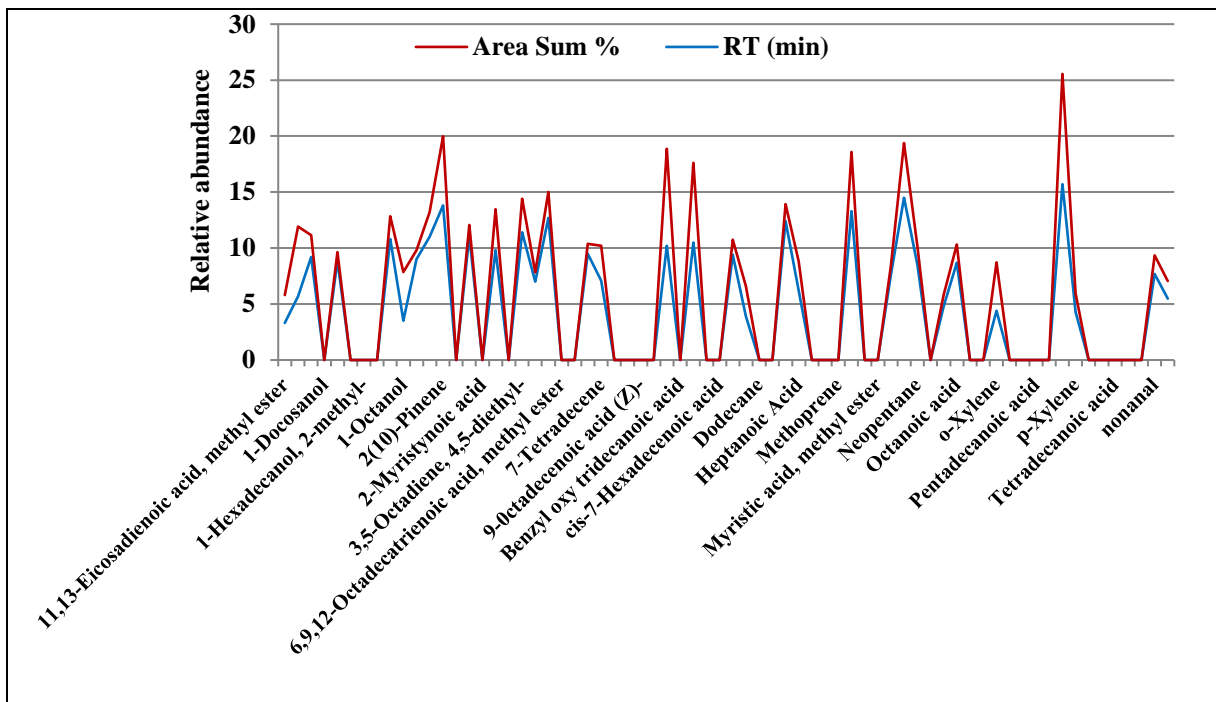


Fig. 3. Chart of GC-mass chromatogram of *Borago Officinalis* L. seed oil from the plants treated with humic acid (4 kg/fed) + Fe (100 ppm) during the second season (2021/2022).

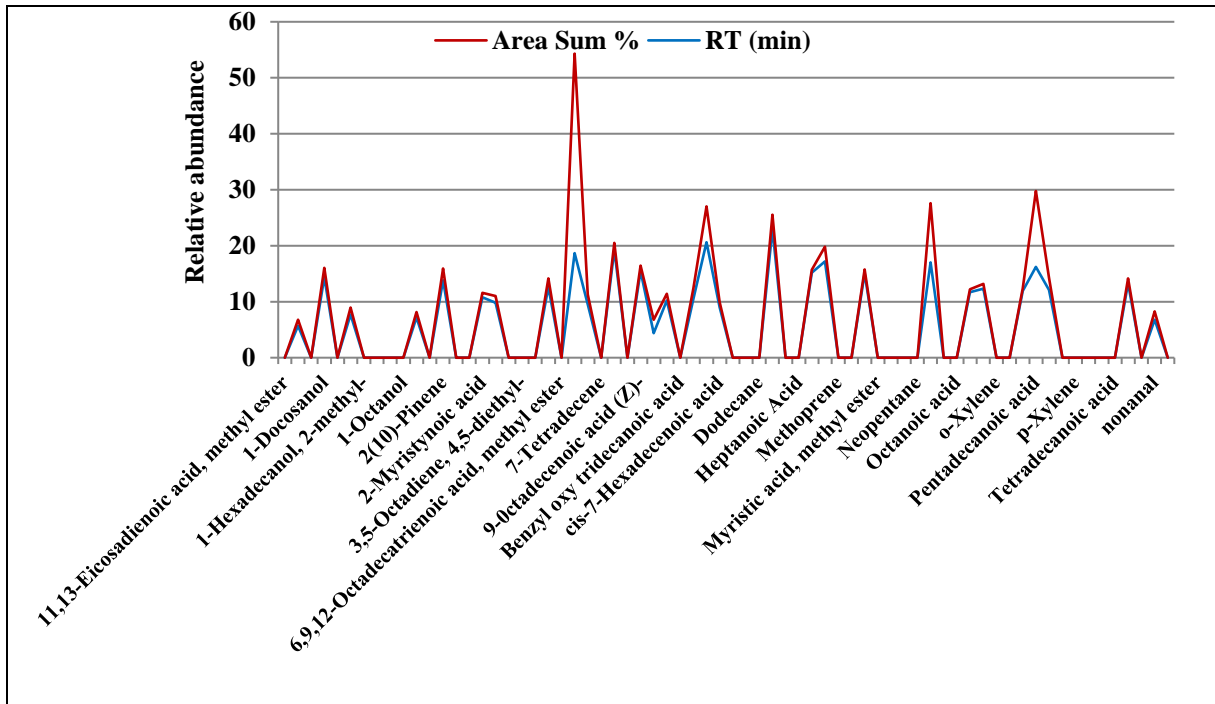


Fig. 4. Chart of GC-mass chromatogram of *Borago Officinalis* L. seed oil from the plants treated with humic acid (4 kg/fed) + Mn (100 ppm) during the second season (2021/2022).

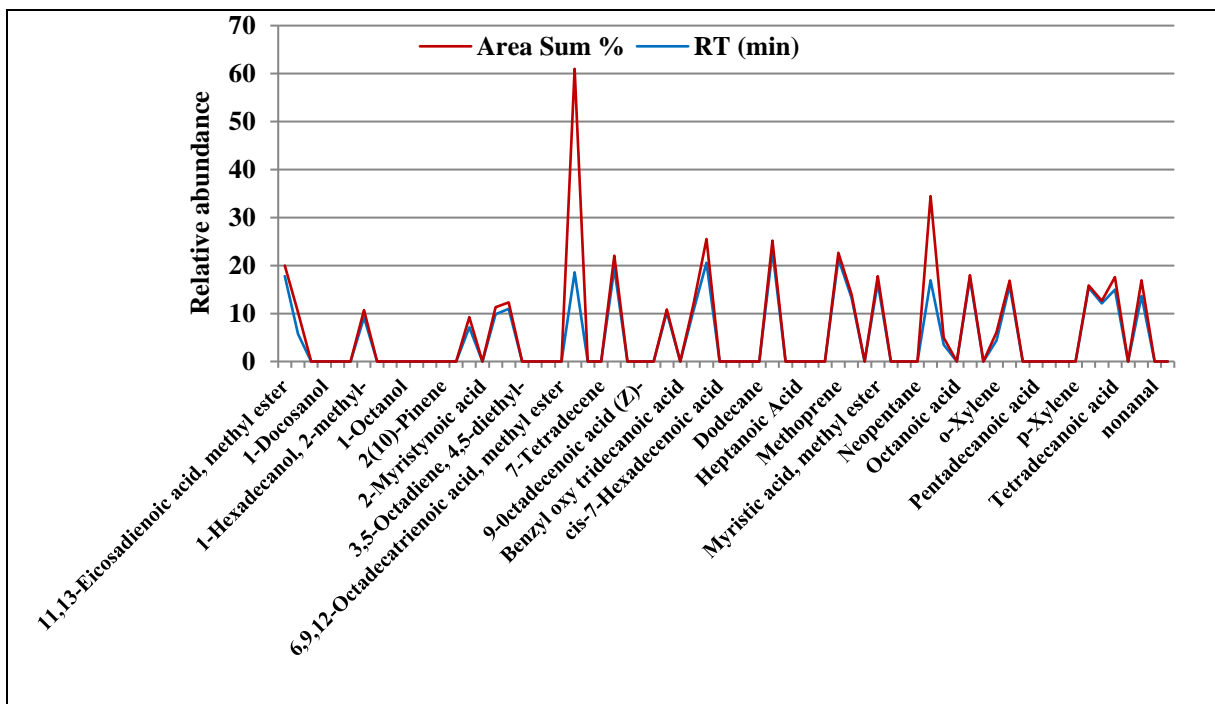


Fig. 5. Chart of GC-mass chromatogram of *Borago Officinalis* L. seed oil from the plants treated with humic acid (4 kg/fed) during the second season (2021/2022).

(1.65%), Methyl γ -linolenate (1.87%), Myristic acid (1.11 %), methyl ester, n-Hexadecanoic acid (16.79 %), Pentadecanoic acid (1.57 %), Tetradecanoic acid (1.8 %) and α -Humulene (5.62%).

While the sample resulted from the treatment of humic acid (4 kg/fed) + Fe (100 ppm) recorded 33 compounds, i.e., 11,13-eicosadienoic acid, methyl ester (2.5%), 1-Decanol (6.23%), 1-Decyne (1.94%), 1-Dodecene (0.74%), 1-Octadecyne (2.03%), 1-Octanol (4.37%), 1-Tetradecanol (0.81%), 1-Undecen-10-al (2.2%), 2 (10)-Pinene (6.17%), 2-Dodecenal, (E)- (0.97%), 2-Undecenal (3.65%), 3,5-Octadiene, 4,5-diethyl- (3.01%), 3-Pinanol (0.84%), 6,8-Dimethoxy-4-methyl-4H-chromene (2.32%), 7-Hexadecyne (0.87%), 7-Tetradecene (3.11%), 9-Octadecyne (8.66%), Camphor (7.11%), Cyclododecene (1.35%), Cyclohexane, 1,1,3-trimethyl (2.7%), nonanal (1.51%), 1-Decene (2.61%), Methyl γ -linolenate (5.28%), Myristoleic acid (0.8%), Myrtenoic acid, butyl ester (4.89%), Neopentane (1.63%), Nonane (1.02%), Octanoic acid (1.63%), o-Xylene (4.33%), Pinolenic acid (9.85%), p-Xylene (1.65%), Heneicosane (1.65%) and Heptanoic Acid (1.57%). The sample resulted from the treatment of humic acid (4 kg/fed) +Mn (100 ppm) recorded 26 compounds, i.e., 1-Decanol (1.1%),1-Docosanol (1.65%),1-Eicosanol (1.25%), 1-Tetradecanol (0.98%), 2(10)-Pinene (2.12%),2-Myristynoic acid (0.79%), 2-Undecenal (1.22%), 6,8-Dimethoxy-4-methyl-4H-chromene (1.56 %), 7,10-Octadecadienoic acid, methyl ester (35.66%), 7-Hexadecyne (1.82%), 8,11,14-Docosatrienoic acid, methyl ester (1.18%), 9-Octadecenoic acid (Z)- (0.94%), 9-Octadecenoic acid (Z)-, phenylmethyl ester (2.38%), 9-Octadecyne (1.22%), Camphor (1.91%), cis-11-Eicosenoic acid (6.44%), cis-7-Hexadecenoic acid (0.94%), Erucic acid (2.55%), Hexadecanoic acid, methyl ester (0.49%), Isopropyl linoleate (2.64%), Myristic acid (0.66%), n-Hexadecanoic acid (10.58%), Oleic Acid (0.52%), Oxiraneoctanoic acid, 3-6ctyl-, cis- (0.91%),

Palmitoleic acid (0.46%), Pentadecanoic acid (13.56%), Phytol (2.15%), Tridecanedial (0.85%) and Nonanal (1.46%).

Furthermore, the sample resulted from the treatment of humic acid (4 kg/fed) recorded 25 compounds, i.e., 11,13-Eicosadienoic acid, methyl ester (2.19 %), 1-Decanol(4.55 %), 1-Hexadecanol, 2-methyl- (1.52 %), 2-Dodecenal, (E)-(2.12 %), 2-Undecenal(1.49 %), 2-Undecenal, E-(1.32 %), 7,10-Octadecadienoic acid, methyl ester(42.41 %), 8,11,14-Docosatrienoic acid, methyl ester(2.65 %), 9-Octadecyne(0.63 %), Camphor(1.38%), cis-11-Eicosenoic acid(4.9 %), Erucic acid (2.13%), Methoprene (1.22%), Methyl γ -linolenate (0.71%), Myristic acid, methyl ester(1.43%), n-Hexadecanoic acid (17.53%), Nonane (1.5%), Oleic Acid (0.65%), o-Xylene (1.73%), Palmitic acid, 2-(tetradecyloxy) ethyl ester (1.07%), Rescinamine (0.47%), Retinal (0.52 %), Tetradecanoic acid (2.63%) and α -Humulene (3.25%). On the other hand, the sample resulted from the control treatment recorded 14 compounds only, i.e., 11,13-Eicosadienoic acid, methyl ester (3.81%), 1-Decanol (3.14%), 1-Decyne (0.5%), 2-Undecenal (1.84%), 7,10-Octadecadienoic acid, methyl ester (49.07%), 8,11,14-Docosatrienoic acid, methyl ester (1.62%), 9-Octadecyne (0.62%), Camphor (1.21%), cis-11-Eicosenoic acid (3.91%), Erucic acid (1.47%), Myristic acid, methyl ester (0.67%), n-Hexadecanoic acid (21.12%), Octanoic acid (2.41%) and Tetradecanoic acid (4.98%).

The main compounds identified in plenty in almost all cases were 11.13-Eicosadienoic acid, methyl ester, 1-Decanol, 2-Undecenal, 9-Octadecyne, Benzyl oxy tridecanoic acid, Camphor and cis-11-Eicosenoic acid. Lastly, only one component viz. 1-Nonyne appeared in the seed oil sample resulting from the treatment of humic acid (4 kg/fed) + Zn (100 ppm).

CONCLUSION

Adding humic acid at 4 kg/fed before sowing as soil application then foliar spraying the plants with Fe or Zn at 100 ppm for 4

times is the best for the growth, flowering characteristics, chemical composition, and oil production of borage plants.

REFERENCES

- Adamczyk-Szabela, D. and Wolf, W.M. (2024). The influence of copper and zinc on photosynthesis and phenolic levels in basil (*Ocimum basilicum* L.), borage (*Borago officinalis* L.), common nettle (*Urtica dioica* L.) and peppermint (*Mentha piperita* L.). *Inter. J. of Mole. Sci.*, 25(7):1-13.
<https://doi.org/10.3390/ijms25073612>
- Aftab, T. and Hakeem, K.R. (2020). Plant micronutrients: deficiency and toxicity management. Springer Nature., Switzerland, 470 p.
<https://doi.org/10.1007/978-3-030-49856-6>.
- Aghdasi, S.; Modarres-Sanavy, S.A.; Agha-Alikhani, M. and Keshavarz, H. (2018). The effect of dehydration and foliar application of Fe and Mn on some physiological and quantitative and qualitative traits of green mung bean forage (*Vigna radiate* L.). *J. Plant Proc. Funct.*, 7:101-115.
- Ahmed, N.; Zhang, B.; Chachar, Z.; Li, J.; Xiao, G.; Wang, Q.; Hayat, F.; Deng, L.; Narejo, M.; Bozdar, B. and Tu, P. (2024). Micronutrients and their effects on horticultural crop quality, productivity and sustainability. *Scientia Horticulturae*, 323:1-9.
<https://doi.org/10.1016/j.scienta.2023.112512>
- Ampong, K.; Thilakaranthna, MS. and Gorim, L.Y. (2022). Understanding the role of humic acids on crop performance and soil health. *Frontiers in Agronomy*, 4:1-10.
<https://doi.org/10.3389/fagro.2022.848621>
- AOAC (1984). *Official Methods of Analysis*. Association of Official Analytical Chemists, Washington DC, USA, 771 p.
- Ariafar, S. and Forouzandeh, M. (2017). Evaluation of humic acid application on biochemical composition and yield of black cumin under limited irrigation condition. *Bull. Soc. R. Sci. de Liège*, 86(1):13-24.
- Asadi-Samani, M.; Bahmani, M. and Rafieian-Kopaei, M. (2014). The chemical composition, botanical characteristic and biological activities of *Borago officinalis*: A review. *Asian Pac. J. Trop. Med.*, 7:22–28.
- Ayobizadeh, N.; Haei, Gh.; Aminidehaghi, M.; Masoud Sinki, J. and Rezvani, Sh. (2019). Effect of foliar application of iron nano-chelate and folic acid on seed yield and some physiological traits of sesame cultivars under drought tension conditions. *Crop Physiol. J.*, 10(40):55-74.
- Azizi, M. and Safaei, Z. (2017). The effect of foliar application of humic acid and nano fertilizer on morphological traits, yield, essential oil content and yield of black cumin (*Nigella sativa* L.). *J. Hort. Sci.*, 30(4):671-680.
- Balafrej, H.; Bogusz, D.; Triqui, Z.E.A.; Guedira, A.; Bendaou, N.; Smouni, A., and Fahr, M. (2020). Zinc hyperaccumulation in plants: A review. *Plants*, 9(5):1-22.
<https://doi.org/10.3390/plants9050562>
- Balandr'an-Valladares, M.I.; Cruz-Alvarez, O.; Jacobo-Cuellar, J.L.; Hernández-Rodríguez, O.A.; Flores-Córdova, M.A.; Parra-Quezada, R.; Sánchez-Chávez, E. and Ojeda-Barrios, D.L. (2021). Changes in nutrient concentration and oxidative metabolism in pecan leaflets at different doses of zinc. *Plant Soil Environ* 67(1):33-39.
- Bastani, S.; Hajiboland, R.; Khatamian, M. and Saket-Oskoui, M. (2018). Nano iron (Fe) complex is an effective source of Fe for tobacco plants grown under low Fe supply. *J. Soil Sci. Plant Nutr.*, 18(2):524–541.

- Bera, B.; Bokado, K. and Arambam, S. (2024). Effect of humic acid on growth, yield and soil properties in rice. *Int. J. Plant Soil Sci.*, 36(6):26-35.
- Black, C.A.; Evans, D.O.; Ensminger, L.E.; White, J.L.; Clark, F.E. and Dinauer, R.C. (1982). *Methods of Soil Analysis, Part 2. Chemical and Microbiological Properties*, 2nd Ed. Soil Science Society of America, Madison, WI, USA, 1159 p.
- Bulgari, R.; Morgutti, S.; Cocetta, G.; Negrini, N.; Farris, S.; Calcante, A.; Spinardi, A.; Ferrari, E.; Mignani, I. and Oberti, R. (2017). Evaluation of borage extracts as potential biostimulant using a phenomic, agronomic, physiological, and biochemical approach. *Frontiers in Plant Science*, 8:1-16.
<https://doi.org/10.3389/fpls.2017.00935>
- Canellas, L.P.; Canellas, N.O.; da S. Irineu, L.E.S.; Olivares, F.L. and Piccolo, A. (2020). Plant chemical priming by humic acids. *Chemical and Biological Technologies in Agriculture*, 7:1-17.
<https://doi.org/10.1186/s40538-020-00178-4>
- Cieschi, M.T.; Polyakov, A.Y.; Lebedev, V.A.; Volkov, D.S.; Pankratov, D.A.; Veligzhanin, A.A.; Perminova, I.V. and Lucena, J.J. (2019). Eco-friendly iron-humic nanofertilizers synthesis for the prevention of iron chlorosis in soybean (*Glycine max*) grown in calcareous soil. *Frontiers Plant Sci.*, 10:1-17.
<https://doi.org/10.3389/fpls.2019.00413>
- Davoodi, S.H.; Biyabani, A.; Rahemi Karizaki, A.; Modares Sanavi, S.A.; Gholamalipour Alamdari, E. and Zaree, M. (2020). Effect of iron and zinc nano chelates on yield and yield components of black cumin medicinal plant (*Nigella sativa*). *Iran. J. Field Crops Res.*, 18(3):267-278.
- Dukpa, P.; Chatterjee, R. and Subba, S.K. (2017). Soil and foliar iron fertilization on terrestrial water spinach (*Ipomoea reptans*) for biofortification. *J. Pharmacognosy and Phytochemistry*, 6(6):1327-1330.
- Duncan, D.B. (1955). Multiple range and multiple F tests. *Biometrics*, 11(1):1-42.
- El Hafid, R.; Blade, S.F. and Hoyano, Y. (2002). Seeding date and nitrogen fertilization effects on the performance of borage (*Borago officinalis* L.). *Industrial Crops and Products*, 16(3):193-199.
- Elankavi, S.; Nambi, J.; Ramesh, S.; Jawahar, S. and Lavanya, K. (2020). Influence of different doses of fertilizers and foliar spray of nutrients on yield and yield attributes of rice. *Annals of the Romanian Society for Cell Biology.*, 24(2):1127-1134.
- El-Gohary, A.E.; El-Sherbeny, S.E.; Ghazal, G.M.; Khalid, K.A. and Hussein, M.S. (2014). Evaluation of essential oil and monoterpenes of peppermint (*Mentha piperita* L.) under humic acid with foliar nutrition. *J. Mater Environ Sci.*, 5(6):1885-1890.
- El-Rahman, A.; Ali, A.F. and Amer, E.H. (2023). Physiological effects of organic and bio fertilizers on borage (*Borago officinalis* L.) plants. *Archives of Agri. Sci. J.*, 6(2):141-164.
- Ezz El-Din, Azza A. and Hendawy, S.F. (2010). Effect of dry yeast and compost tea on growth and oil content of *Borago officinalis* plant. *Res J. Agric. Biol. Sci.*, 6(4):424-430.
- Galambosi, B.; Domokos, J. and Sairanen, J. (2014). Experiences with different methods of harvesting borage (*Borago officinalis*). *Z. Für Arznei Gewürz Pflanz.*, 19:61-66.
- Ghorbani, P.; Eshghi, S.; Ershadi, A.; Shekafandeh, A. and Razzaghi, F. (2019). The possible role of foliar application of manganese sulfate on mitigating adverse effects of water stress in grapevine. *Commun Soil Sci. Plant Anal.*, 50(13):1550-1562

- Gilbertson, P.K.; Berti, M.T. and Johnson, B.L. (2014). Borage cardinal germination temperatures and seed development. *Ind. Crops Prod.*, 59:202–209.
- Gomes, D.G.; Pieretti, J.C.; Rolim, W.R.; Seabra, A.B. and Oliveira, H.C. (2020). Advances in nano-based delivery systems of micronutrients for a greener agriculture. In: Jogaiah, S.; Singh, H.B.; Fraceto, L.F. and Lima, R. (eds.), *Advances in Nano-Fertilizers and Nano-Pesticides in Agriculture: A Smart Delivery System for Crop Improvement*, Woodhead Publishing, Elsevier Inc., USA, pp. 111-143.
<https://doi.org/10.1016/C2019-0-01205-8>
- Gupta, M. and Singh, S. (2010). *Borago officinalis* L.; An important medicinal plant of Mediterranean region: A review. *Int. J. Pharm. Sci. Rev. Res.*, 5(1):27–34.
- Hayati, A.; Rahimi, M.M.; Kledari, A. and Hosseini, S.M. (2022). Impact of humic acid and nano-Fe chelate on improving vegetative traits, yield and essential oil content of black cumin (*Nigella sativa*) under drought stress. *Iranian J. of Horti. Sci. and Tech.*, 23(1):179-190.
- Herbert, D.; Phipps, P.J. and Strange, R.E. (1971). Determination of total carbohydrates, *Methods in Microbiology*, 5(8):290-344.
- Herlihy, J.H.; Long, T.A. and McDowell, J.M. (2020). Iron homeostasis and plant immune responses: recent insights and translational implications. *J. Biol. Chem.*, 295(39):13444-13457.
- Horneck, D.A. and Hanson, D. (1998). Determination of potassium and sodium by flame Emission spectrophotometry. In: Kalra, Y.P. (ed.), *Handbook of Reference Methods for Plant Analysis*, CRC Press, USA, pp.153-155.
- Horneck, D.A. and Miller, R.O. (1998). Determination of total nitrogen in plant. In: Kalra, Y.P. (ed.), *Handbook of Reference Methods for Plant Analysis*, CRC Press, USA, pp. 75-83.
- Hoseini, M.; Paknejad, F. and Ilkaee, M.N. (2023). Evaluation of humic acid and iron and zinc nanochelates effect on Italian basil (*Ocimum basilicum* L.) in salinity stress condition. *J. of Org. Far. of Med. Plants*, 2(1):44-51.
- Hucker, T. and Catroux, G. (1980). Phosphorus in sewage ridge and animal's wastes slurries. *Proc. the EEC Seminar, Haren (Gr), The Netherlands*, 443 p.
- Jackson, M.L. (1973). *Soil Chemical Analysis*; Prentice-Hall of Indian Private: New Delhi, India, 498 p.
- Jindo, K.; Canellas, L.P.; Albacete, A.; Figueiredo dos Santos, L.; Frinhan Rocha, R.L.; Carvalho Baia, D.; Canellas, N.O.A.; Goron, T.L. and Olivares, F.L. (2020). Interaction between humic substances and plant hormones for phosphorous acquisition. *Agronomy*, 10(5):1-18.
<https://doi.org/10.3390/agronomy10050640>
- Kazemi, S.; Pirmoradi, M.R.; Karimi, H.; Raghani, M.; Rahimi, A.; Kheiry, A. and Malekzadeh, M.R. (2023). Effect of foliar application of humic acid and zinc sulfate on vegetative, physiological, and biochemical characteristics of *Physalis alkekengi* L. under soilless culture. *J. of Soil Sci. and Plant Nutri.*, 23(3):3845-3856.
- Khaled, H. and Fawy, H.A. (2011). Effect of different levels of humic acids on the nutrient content, plant growth, and soil properties under conditions of salinity. *Soil and Water Research*, 6(1):21-29.
- Khosravi, F.; Bahmanyar, M.A. and Akbarpour, V. (2023). Effect of different levels of humic acid and zinc sulfate on morphological and phytochemical traits of (*Salvia officinalis* L.). *J. of Horti. Sci.*, 37(3):615-627.
- Korani, M.; Ilkaee, M.N. and Paknejad, F. (2023). Foliar application of humic acid and iron and zinc nano chelates alleviate the salinity damage on chicory

- (*Cichorium intybus* L.) in hydroponic culture. *J. of Org. Farm. of Med. Pl.*, 2(1):1-8.
- Li, J.; Cao, X.; Jia, X.; Liu, L.; Cao, H.; Qin, W. and Li, M. (2021). Iron deficiency leads to chlorosis through impacting chlorophyll synthesis and nitrogen metabolism in areca catechu L. *Front. Plant Sci.*, 12:1-17.
<https://doi.org/10.3389/fpls.2021.710093>
- Memon, S.A.; Baloch, R.A. and Baloch, M.H. (2014). Influence of humic acid and micronutrients (zinc+ manganese) application on growth and yield of phlox (*Phlox paniculata*). *J. of Agri. Tech. Vol.* 10(6):1531-1543.
- Mohamed, Y. and Ghatas, Y. (2020). Effect of some safety growth stimulants and zinc treatments on growth, seeds yield, chemical constituents, oil productivity and fixed oil constituents of chia (*Salvia hispanica* L.) plant. *Sci. J. of Flowers and Ornamental Plants*, 7(2):163-183.
- Montaner, C.; Zufiaurre, R.; Movila, M.; Mallor, C. (2022). Evaluation of borage (*Borago officinalis* L.) genotypes for nutraceutical value based on leaves fatty acids composition. *Foods*, 11(1):1-19.
<https://doi.org/10.3390/foods11010016>
- Mubarak, D.M.; Yasser, A.; El-Azab, M.E. and Aly, M.M. (2023). Impact of spraying chelated zinc with humic acid extracted from compost on the growth, yield and nutritional status of japanese cabbage plants grown in sandy soil. *Euro. J. of Bio. Sci.*, 15(1):1-7.
- Nardi, S.; Schiavon, M. and Francioso, O. (2021). Chemical structure and biological activity of humic substances define their role as plant growth promoters. *Molecules.*, 26(8):1-20.
<https://doi.org/10.3390/molecules26082256>
- Omar, A.A. (2020). Response of Caraway Plant to Zinc and Humic Acid Treatments. M.Sc. Thesis, Fac. Agric., Assiut Univ., Egypt, 203 p.
- Otiende, M.A.; Fricke, K.; Nyabundi, J.O.; Ngamau, K.; Hajirezaei, M.R. and Druge, U. (2021). Involvement of the auxin–cytokinin homeostasis in adventitious root formation of rose cuttings as affected by their nodal position in the stock plant. *Planta*, 254(4):1-17.
<https://doi.org/10.1007/s00425-021-03709-x>
- Russo, R.O. and Berlyn, G. (1990). The use organic biostimulants to help low input sustainable agriculture. *J. Sust. Agric.*, 1:19-42.
- Seifzadeh, A.R.; Khaledian, M.R.; Zavareh, M.; Shahinrokhsar, P. and Damalas, C.A. (2020). European borage (*Borago officinalis* L.) yield and profitability under different irrigation systems. *Agriculture*, 10(4):1-13.
<https://doi.org/10.3390/agriculture10040136>
- Shah, ZH.; Rehman, HM.; Akhtar, T.; Alsamadany, H.; Hamooh, BT.; Mujtaba, T.; Daur, I.; Al Zahrani, Y.; Alzahrani, HA.; Ali, S. and Yang, SH. (2018). Humic substances: Determining potential molecular regulatory processes in plants. *Frontiers in Plant Science*, 9:1-12.
<https://doi.org/10.3389/fpls.2018.00263>
- Sheikhzadeh, P.; Zare, N. and Mahmoudi, F. (2021). The synergistic effects of hydro and hormone priming on seed germination, antioxidant activity and cadmium tolerance in borage. *Acta Bot. Croat.*, 80(1):18–28.
- Şimşek, O. and Çelik, H. (2021). Effects of iron fortification on growth and nutrient amounts of spinach (*Spinacia oleracea* L.). *J. Plant Nutr.*, 44(18):2770–2782.
- Snedecor, G.W. and Cochran, W.G. (1989). *Statistical Methods*, 8th Ed.; Iowa State University Press: Ames, USA., 54:71-82
- Suganya, A.; Saravanan, A. and Manivannan, N. (2020). Role of zinc nutrition for increasing zinc availability, uptake, yield, and quality of maize (*Zea Mays* L.) grains:

- an overview. *Commun. Soil Sci. Plant Anal.*, 51(15):2001-2021.
- Tawfik, O.H. (2022). Influence of zinc and humic acid on growth, yield and essential oil percentage of fennel (*Foeniculum vulgare* Mill.) plants. *Sci. J. of Flowers and Ornamental Plants*, 9(4):397-407.
- Tripathi, R.; Tewari, R.; Singh, K.P.; Keswani, C.; Minkina, T.; Srivastava, A.K.; De Corato, U. and Sansinenea, E. (2022). Plant mineral nutrition and disease resistance: a significant linkage for sustainable crop protection. *Front. Plant Sci.*, 13:1-12.
<https://doi.org/10.3389/fpls.2022.883970>
- Turan, M.; Ekinci, M.; Kul, R.; Kocaman, A.; Argin, S.; Zhirkova, A.M.; Perminova, I.V. and Yildirim, E. (2022). Foliar applications of humic substances together with Fe/nano Fe to increase the iron content and growth parameters of spinach (*Spinacia oleracea* L.). *Agro.*, 12(9):1-19.
<https://doi.org/10.3390/agronomy12092044>
- Vafa, Z.N.; Sohrabi, Y. and Samir, Z. (2020). Humic acid and nano Zn chelated fertilizer regulates nutrient uptake and growth and production of summer savory. *J. of Bio. and Nat.*, 11(4):5-15.
- Wu, S.; Li, R.; Peng, S.; Liu, Q. and Zhu, X. (2017). Effect of humic acid on transformation of soil heavy metals. In *IOP Conference Series: Materials Science and Engineering.*, 207(1):1-7.
<https://doi.org/10.1088/1757-899X/207/1/012089>
- Yadegari, M. (2023). The effects of foliar application of some micronutrient elements on the content and composition of the essential oil of damask rose (*Rosa damascena* Mill.). *Iran Agri. Res.*, 41(2)35-49.
- Ye, Y.; Medina-Velo, I.A.; Cota-Ruiz, K.; Moreno-Olivas, F. and Gardea-Torresdey, J.L. (2019). Can abiotic stresses in plants be alleviated by manganese nanoparticles or compounds? *Ecotoxicol. Environ. Saf.*, 184:1-37.
<https://doi.org/10.1016/j.ecoenv.2019.109671>
- Yosef, A.H.E.; Soliman, Y. and Elsayed, A. (2024). Effect of planting date and foliar application of humic acid on vegetative growth, yield, and fixed oil of black cumin plants. *Scientific Journal of Flowers and Ornamental Plants*, 11(1):17-26.
- Zghair, A.A.; Hassan, F.A. and Jrry, A.N. (2022). Study the effect of spraying with humic acid and zinc on the yield of the volatile oil of *Rosmarinus officinalis* L. and its physical properties. *Biochem. Cell. Arch.*, 22(1):2727-2731.

تأثير حمض الهيوميك وبعض العناصر الغذائية الصغرى على نمو وإنتاجية البذور ومحتوى الزيت في نبات خبز النحل

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يعتبر خبز النحل (*Borago officinalis* L.) نباتاً عشبياً حولياً ينتمي إلى الفصيلة البوراجينية ويعرف باسم نبات النحل وهو نبات طبي مهم أصلي في منطقة البحر الأبيض المتوسط وانتشر على نطاق واسع في العديد من البلدان الأخرى. أجريت تجربة بهدف تقييم تأثير حمض الهيوميك والعناصر الغذائية الصغرى على خصائص النمو والتركيب الكيميائي وإنتاج الزيت في نباتات خبز النحل. أجريت في المزرعة التجريبية لقسم البساتين، كلية الزراعة، مشتهر، جامعة بنها، خلال موسم النمو المتتاليين ٢٠٢٠/٢٠٢١ و ٢٠٢١/٢٠٢٢. تضمنت هذه التجربة ١٦ معاملة ناتجة عن التفاعل بين إضافة حمض الهيوميك للتربة بمعدل صفر (الكنترول) و ٢ و ٣ و ٤ كجم/فدان والرشي الورقي بالماء (الكنترول) أو الزنك أو الحديد أو المنجنيز بمعدل ١٠٠ جزء في المليون لكل منها. أشارت النتائج إلى أن معاملة حمض الهيوميك بمعدل ٤ كجم/فدان + حديد بمعدل ١٠٠ جزء في المليون أدت إلى تحسين كبير في خصائص النمو بما في ذلك عدد الفروع والوزن الطازج والجاف للنبات. كما أنتجت معاملة حمض الهيوميك بمعدل ٤ كجم/فدان + حديد بمعدل ١٠٠ جزء في المليون أقصى قيم للنيتروجين الكلي والفوسفور والبوتاسيوم والكربوهيدرات في كلا الموسمين. وفي الوقت نفسه، أدت معاملة حمض الهيوميك بمعدل ٤

كجم/فدان + زنك بمعدل ١٠٠ جزء في المليون إلى تحسين كبير في بعض الصفات بما في ذلك طول النبات والوزن الطازج والجاف للنورات وإنتاج البذور في كلا الموسمين. أعطت معاملة حمض الهيوميك بمعدل ٤ كجم/فدان + زنك بمعدل ١٠٠ جزء في المليون أعلى نسبة ثابتة للزيت يليها معاملة حمض الهيوميك بمعدل ٤ كجم/فدان + حديد بمعدل ١٠٠ جزء في المليون. تم ظهور ٦٨ مركباً في مكونات عينات زيت البذور أثناء تحليل العينات على جهاز GC-MS. حيث سجلت العينة الناتجة عن معاملة حمض الهيوميك (٤ كجم/فدان) + حديد (١٠٠ جزء في المليون) ٣٣ مركباً، ومن ناحية أخرى سجلت العينة الناتجة عن معاملة الكنترول ١٤ مركباً.